

18MBU601A
(4H – 4C)

VI
MUSHROOM CULTIVATION

Instruction Hours / week:L: 0 T: 0 P: 4
External: 60 Total: 100

Marks: Internal: 40

End Semester Exam: 3 Hours

COURSE OBJECTIVES

To teach on classification, cultivation, diseases and health benefits of mushrooms.

COURSE OUTCOME

Provides knowledge on various mushrooms and its cultivation techniques

Unit I

Mushroom morphology: Different parts of a typical mushroom and variations in mushroom morphology. Key to differentiate edible from poisonous mushrooms. Mushroom Classification: Based on occurrence – Epigenous and hypogenous, Natural habitats – Humicolous, Lignicolous & Coprophilous, Color of spores – white, yellow, pink, purple brown and black. Ainsworth et al classification (8th edition) and Bisby's 'Dictionary of Fungi'.

Unit II

Biology of Mushrooms: Vegetative characters, general morphology, spore germination and life cycle of button mushroom (*Agaricus bisporus*), milky mushroom (*Calocybe indica*), oyster mushroom (*Pleurotus sajorcaju*) and paddy straw mushroom (*Volvariella volvcea*).

Unit III

Equipment and sterilization techniques. Isolation and culture of spores, culture media preparation. Production of mother spawn, multiplication of spawn – Inoculation technique – Cultivation technology – Substrates, composting technology, bed, polythene bag preparation, spawning – casing – cropping – Mushroom production – harvest – packing, storage and marketing.

Unit IV

Nutritional profile of Mushrooms: protein, amino acids, calorific values, carbohydrates, fats, vitamins & minerals. Medicinal Properties of Mushrooms: Antibacterial, antifungal, antiviral, anti-tumour effect and hematological value. Cardiovascular and renal effect, in therapeutic diets, adolescence, for aged persons and diabetes mellitus. Mushroom nutraceuticals.

Unit V

Problems in cultivation – diseases, pests and nematodes, weed moulds and their management strategies. Mushroom economics: economics of spawn and mushroom, cultivation, postharvest technologies. Processing and preservation of mushrooms. Mushroom research centres in India.

SUGGESTED READINGS

1. Alice, D., Muthusamy and Yesuraja, M. (1999). Mushroom Culture. Agricultural College, Research Institute Publications, Madurai.

2. Marimuthu, T. et al. (1991). Oyster Mushroom. Department of Plant Pathology. Tamil Nadu Agricultural University, Coimbatore.
3. Nita Bhal. (2000). Handbook on Mushrooms. 2nd ed. Vol. I and II. Oxford and IBH Publishing Co. Pvt. Ltd., New Delhi.
4. Pathak, V. N. and Yadav, N. (1998). Mushroom Production and Processing Technology. Agrobios, Jodhpur.
5. Tewari Pankaj Kapoor, S. C. (1988). Mushroom Cultivation. Mittal Publication, New Delhi.

LECTURE PLAN-UNIT-1

S.NO	Lecture duration hour	Topics	Supporting materials
1	1	Introduction	T1-1-8
2	1	Mushroom morphology	T1 55-59
3	1	Different parts of a typical mushroom and variations in mushroom morphology. Key to differentiate edible from poisonous mushrooms	T1 118-119
4	1	Mushroom Classification: Based on occurrence	T1-59-61
5	1	classification-natural habitats	T1 61
6	1	classification-based n coloured spores	T1 61-70
7	2	Ainsworth et al classification (8th edition) and Bisby's 'Dictionary of Fungi'. Ainsworth et al classification (8th edition) and Bisby's 'Dictionary of Fungi'.	T1 85-89
8	1	Unit Revision	
Textbooks:		T1-mushroom cultivation-D.P.Tripathi	
Journals:		-	
Website:		-	
Reference books:			
Total number of hours		9	

LECTURE PLAN-UNIT-2			
S.NO	Lecture duration hour	Topics	Supporting materials
1	2	Life cycle of button mushroom	T1-181-190
2	2	life cycle of milky mushroom	T1 255-256
3	2	life cycle of oyster mushroom	T 257
4	2	life cycle of paddy straw mushroom	T1 258-260
5	1	Unit Revision	
Textbooks:		T1-mushroom cultivation-D.P.Tripathi	
Journals:		-	
Website:		-	
Reference books:			
Total number of hours		9	

LECTURE PLAN-UNIT-3			
S.NO	Lecture duration hour	Topics	Supporting materials
1	1	Equipment and sterilization techniques	T1-125-129
2	1	Isolation and culture of spores, culture media preparation	T1 160-167
3	1	Production of mother spawn, multiplication of spawn – Inoculation technique	T1 156-160
4	1	Cultivation technology – Substrates, composting technology, bed, polythene bag preparation,	T1-137-153
5	1	spawning – casing – cropping – Mushroom production – harvest	T1 154-156
6	1	packing, storage and marketing	T1-298-305
7	1	unit revision	
Textbooks:		T1-mushroom cultivation-D.P.Tripathi	
Journals:		-	
		-	
Website:			
Reference books:			
Total number of hours		7	

LECTURE PLAN-UNIT-4

S.NO	Lecture duration hour	Topics	Supporting materials
1	1	Nutritional profile of Mushrooms: protein, amino acids	T1-25-27
2	1	Nutritional profile-calorific values, carbohydrates, fats, vitamins & minerals	T1 28-34
3	1	Medicinal Properties of Mushrooms: Antibacterial, antifungal	T1 36
4	1	antiviral, anti-tumour effect and hematological value	T1-37
5	1	Cardiovascular and renal effect,	T1 38
6	1	in therapeutic diets, adolescence, for aged persons and diabetes mellitus.	T1-39-41
7	1	Mushroom nutraceuticals.	
Textbooks:		T1-mushroom cultivation-D.P.Tripathi	
Journals:		-	
Website:		-	
Reference books:			
Total number of hours		7	

LECTURE PLAN UNIT-5

S.NO	Lecture duration hour	Topics	Supporting materials
1	1	Problems in cultivation – diseases, pests	T1-284-293
2	1	Problems in cultivation – nematodes, weed moulds and their management strategies	T1 293-296
3	1	Mushroom economics: economics of spawn and mushroom	T1 25
4	1	cultivation, postharvest technologies	T1-298-301
5	2	Processing and preservation of mushrooms.	T1 301-305
6	1	Mushroom research centres in India.	T1-306
7	1	unit revision	
Textbooks:		T1-mushroom cultivation-D.P.Tripathi	
Journals:		-	
Website:		-	
Reference books:			
Total number of hours		8	

Unit I

Unit I

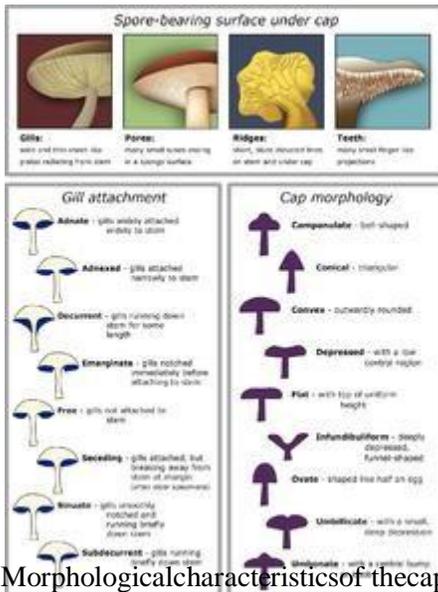
Mushroom morphology: Different parts of a typical mushroom and variations in mushroom morphology. Key to differentiate edible from poisonous mushrooms. Mushroom Classification: Based on occurrence—Epigenous and hypogeous, Natural habitats—Humicolous, Lignicolous & Coprophilous, Color of spores—white, yellow, pink, purple brown and black. Ainsworth et al classification (8th edition) and Bisby's 'Dictionary of Fungi'.

A **mushroom**, or **toadstool**, is the fleshy, spore-bearing fruiting body of a fungus, typically produced above ground on soil or on its food source.

The standard for the name "mushroom" is the cultivated white button mushroom, *Agaricus bisporus*; hence the word "mushroom" is most often applied to those fungi (Basidiomycota, Agaricomycetes) that have a stem (stipe), a cap (pileus), and gills (lamellae, sing. lamella) on the underside of the cap. "Mushroom" also describes a variety of other gilled fungi, with or without stems, therefore the term is used to describe the fleshy fruiting bodies of some Ascomycota. These gills produce microscopic spores that help the fungus spread across the ground or its occupant surface.

Forms deviating from the standard morphology usually have more specific names, such as "bolete", "puffball", "stinkhorn", and "morel", and gilled mushrooms themselves are often called "agarics" in reference to their similarity to *Agaricus* or their order Agaricales. By extension, the term "mushroom" can also designate the entire fungus when in culture; the thallus (called mycelium) of species forming the fruiting bodies called mushrooms; or the species itself.

Identification



Morphological characteristics of the caps of mushrooms

Identifying mushrooms requires a basic understanding of their macroscopic structure. Most are Basidiomycetes and gilled. Their spores, called basidiospores, are produced on the gills and fall in a fine rain of powder from under

the caps as a result. At the microscopic level, the basidiospores are shot off basidia and then fall between the gills in the dead air space. As a result, for most mushrooms, if the caps are cut off and placed gill-side-down overnight, a powdery impression reflecting the shape of the gills (or pores, or spines, etc.) is formed (when the fruit body is sporulating). The color of the powdery print, called a spore print, is used to help classify mushrooms and can help to identify them. Spore print colors include white (most common), brown, black, purple-brown, pink, yellow, and creamy, but almost never blue, green, or red.^[1]

While modern identification of mushrooms is quickly becoming molecular, the standard methods for identification are still used by most and have developed into a fine art harking back to medieval times and the Victorian era, combined with microscopic examination. The presence of juices upon breaking, bruising reactions, odors, tastes, shades of color, habitat, habit, and season are all considered by both amateur and professional mycologists. Tasting and smelling mushrooms carries its own hazards because of poisons and allergens. Chemical tests are also used for some genera.^[2]

In general, identification to genus can often be accomplished in the field using a local mushroom guide. Identification to species, however, requires more effort; one must remember that a mushroom develops from a button stage into a mature structure, and only the latter can provide certain characteristics needed for the identification of the species. However, over-mature specimens lose features and cease producing spores. Many novices have mistaken humid water marks on paper for white spore prints, or discolored paper from oozing liquids on lamella edges for colored spore prints.

Classification

Main articles: Sporocarp (fungi), Basidiocarp, and Ascocarp



A mushroom (probably *Russula brevipes*) parasitized by *Hypomyces lactifluorum* resulting in a "Lobster mushroom".

Typical mushrooms are the fruit bodies of members of the order Agaricales, whose type genus is *Agaricus* and type species is the field mushroom, *Agaricus campestris*. However, in modern molecularly defined classifications, not all members of the order Agaricales produce mushroom fruit bodies, and many other gilled fungi, collectively called mushrooms, occur in other orders of the class Agaricomycetes. For example, chanterelles are in the Cantharellales, false chanterelles such as *Gomphus* are in the Gomphales, milk-caps mushrooms (*Lactarius*, *Lactifluus*) and russulas (*Russula*), as well as *Lentinellus*, are in the Russulales, while the tough, leathery genera *Lentinus* and *Panus* are among the Polyporales, but *Neolentinus* is in the Gloeophyllales, and the little pin-mushroom genus, *Rickenella*, along with similar genera, are in the Hymenochaetales.

Within the main body of mushrooms, in the Agaricales, are common fungi like the common fairy-ring mushroom, shiitake, enoki, oyster mushrooms, fly agarics and other Amanitas, magic mushrooms like species of *Psilocybe*, paddy straw mushrooms, shaggymanes, etc.

An atypical mushroom is the lobster mushroom, which is a deformed, cooked-lobster-colored parasitized fruit body of a *Russula* or *Lactarius*, colored and deformed by the mycoparasitic Ascomycete *Hypomyces lactifluorum*.^[3]

Other mushrooms are not gilled, so the term "mushroom" is loosely used, and giving a full account of their classifications is difficult. Some have pores underneath (and are usually called boletes), others have spines, such as the hedgehog mushroom and other tooth fungi, and so on. "Mushroom" has been used for polypores, puffballs, jelly fungi, coral fungi, bracket fungi, stinkhorns, and cup fungi. Thus, the term is more one of common application to macroscopic fungal fruiting bodies than one having precise taxonomic meaning. Approximately 14,000 species of mushrooms are described.^[4]

Etymology



Amanita muscaria, the most easily recognised "toadstool", is frequently depicted in fairy stories and on greeting cards. It is often associated with gnomes.^[5]

The terms "mushroom" and "toadstool" go back centuries and were never precisely defined, nor was there consensus on application. Between 1400 and 1600 AD, the terms *mushrom*, *mushrum*, *muscheron*, *moussheroms*, *mussheron*, or *musseroun* were used.^[6]

The term "mushroom" and its variations may have been derived from the French word *mousseron* in reference to moss (*mousse*). Delineation between edible and poisonous fungi is not clear-cut, so a "mushroom" may be edible, poisonous, or unpalatable.

Cultural or social phobias of mushrooms and fungus may be related. The term "fungophobia" was coined by William Delisle Hay of England, who noted a national superstition or fear of "toadstools".^{[7][8][9]}

The word "toadstool" has apparent analogies in Dutch *padde(n)stoel* (toad-stool/chair, mushroom) and German *Krötenschwamm* (toad-fungus, alt. word for panther cap). In German folklore and old fairy tales, toads are often depicted sitting on toadstool mushrooms and catching, with their tongues, the flies that are said to be drawn to the *Fliegenpilz*, a German name for the toadstool, meaning "flies' mushroom". This shows the mushroom got another of its names, *Krötenstuhl* (a less-used German name for the mushroom), literally translating to "toad-stool".

Morphology



Amanita jacksonii buttons emerging from their universal veils



The blue gills of *Lactarius indigo*, a milk-cap mushroom

A mushroom develops from a nodule, or pinhead, less than two millimeters in diameter, called a primordium, which is typically found on or near the surface of the substrate. It is formed within the mycelium, the mass of threadlike hyphae that make up the fungus. The primordium enlarges into a roundish structure of interwoven hyphae roughly resembling an egg, called a "button". The button has a cottony roll of mycelium, the universal veil, that surrounds the developing fruit body. As the egg expands, the universal veil ruptures and may remain as a cup, or volva, at the base of the stalk, or as warts or volval patches on the cap. Many mushrooms lack a universal veil, therefore they do not have either a volva or volval patches. Often, a second layer of tissue, the partial veil, covers the blade-like gill that bears spores. As the cap expands, the veil breaks, and remnants of the partial veil may remain as a ring, or annulus, around the middle of the stalk or as fragments hanging from the margin of the cap. The ring may be skirt-like as in some species of *Amanita*, collar-like as in many species of *Lepiota*, or merely the faint remnants of a cortina (a partial veil composed of filaments resembling a spiderweb), which is typical of the genus *Cortinarius*. Mushrooms lacking partial veils do not form an annulus.^[10]

The stalk (also called the stipe, or stem) may be central and support the cap in the middle, or it may be off-center and/or lateral, as in species of *Pleurotus* and *Panus*. In other mushrooms, a stalk may be absent, as in the polypores that form shelf-like brackets. Puffballs lack a stalk, but may have a supporting base. Other mushrooms, such as truffles, jellies, earthstars, and bird's nests, usually do not have stalks, and a specialized mycological vocabulary exists to describe their parts.

The way the gills attach to the top of the stalk is an important feature of mushroom morphology. Mushrooms in the genera *Agaricus*, *Amanita*, *Lepiota* and *Pluteus*, among others, have free gills that do not extend to the top of the stalk. Others have decurrent gills that extend down the stalk, as in the genera *Omphalotus* and *Pleurotus*. There are a great number of variations between the extremes of free and decurrent, collectively called attached gills. Finer distinctions are often made to distinguish the types of attached gills: adnate gills, which adjoin squarely to the stalk; notched gills, which are notched where they join the top of the stalk; and nexed gills, which curve upward to meet the stalk, and so on. These distinctions between attached gills are sometimes difficult to interpret, since gill attachment may change as the mushroom matures, or with different environmental conditions.^[11]

Microscopic features



[Morchella elata](#) asci viewed with phase contrast microscopy

A hymenium is a layer of microscopic spore-bearing cells that covers the surface of gills. In the nongilled mushrooms, the hymenium lines the innersurfaces of the tubes of boletes and polypores, or covers the teeth of spinefungi and the branches of corals. In the Ascomycota, spores develop within microscopic elongated, sac-like cells called asci, which typically contain eight spores in each ascus. The Discomycetes, which contain the cup, sponge, brain, and some club-like fungi, develop an exposed layer of asci, as on the innersurfaces of cupfungi or within the pits of morels. The Pyrenomycetes, tiny dark-colored fungi that live on a wide range of substrates including soil, dung, leaf litter, and decaying wood, as well as other fungi, produce minute, flask-shaped structures called perithecia, within which the asci develop.^[12]

In the Basidiomycetes, usually four spores develop on the tips of thin projections called sterigmata, which extend from club-shaped cells called a basidia. The fertile portion of the Gasteromycetes, called a gleba, may become powdery as in the puffballs or slimy as in the stinkhorns. Interspersed among the asci are thread-like sterile cells called paraphyses. Similar structures called cystidia often occur within the hymenium of the Basidiomycota. Many types of cystidia exist, and assessing their presence, shape, and size is often used to verify the identification of a mushroom.^[12]

The most important microscopic feature for identification of mushrooms is the spores. Their color, shape, size, attachment, ornamentation, and reaction to chemical tests often can be the crux of an identification. A spore often has a protrusion at one end, called an apiculus, which is the point of attachment to the basidium, termed the apical germ pore, from which the hypha emerges when the spore germinates.^[12]

Growth



Agaricus bitorquus mushroom emerging through asphalt concrete in summer

Many species of mushrooms seemingly appear overnight, growing or expanding rapidly. This phenomenon is the source of several common expressions in the English language including "to mushroom" or "mushrooming" (expanding rapidly in size or scope) and "to pop up like a mushroom" (to appear unexpectedly and quickly). In reality all species of mushrooms take several days to form primordial mushroom fruit bodies, though they do expand rapidly by the absorption of fluids.

The cultivated mushroom as well as the common field mushroom initially form a minute fruiting body, referred to as the pin stage because of their small size. Slightly expanded they are called buttons, once again because of the relative size and shape. Once such stages are formed, the mushroom can rapidly pull in water from its mycelium and expand, mainly by inflating preformed cells that took several days to form in the primordia.

Similarly, there are even more ephemeral mushrooms, like *Parasola plicatilis* (formerly *Coprinus plicatilis*), that literally appear overnight and may disappear by late afternoon on a hot day after rain fall.^[13] The primordia form at ground level in lawns in humid spaces under the thatch and after heavy rain fall or in dewy conditions balloon to full size in a few hours, release spores, and then collapse. They "mushroom" to full size.

Not all mushrooms expand overnight; some grow very slowly and add tissue to their fruit bodies by growing from the edges of the colony or by inserting hyphae. For example, *Pleurotus nebrodensis* grows slowly, and because of this combined with human collection, it is now critically endangered.^[14]



Yellow flowerpot mushrooms (*Leucocoprinus birnbaumii*) at various states of development

Though mushroom fruiting bodies are short-lived, the underlying mycelium can itself be long-lived and massive. A colony of *Armillaria solidipes* (formerly known as *Armillaria ostoyae*) in Malheur National Forest in the United States is estimated to be 2,400 years old, possibly older, and spans an estimated 2,200 acres (8.9 km²). Most of the fungus is underground and in decaying wood or dying tree roots in the form of white mycelia combined with black shoelace-like rhizomorphs that bridge colonized separated wood substrates.^[15]

It has been suggested that the electrical stimulus of a lightning bolt striking mycelia in logs accelerates the production of mushrooms.^[16]

Edible mushrooms

Main articles: [Edible mushroom](#), [Mushroom hunting](#), and [Fungiculture](#)



The *Agaricus bisporus*, one of the most widely cultivated and popular mushrooms in the world



Culinary mushrooms are available in a wide diversity of shapes and colors at this market stand at the San Francisco Ferry Building

Mushrooms are used extensively in cooking, in many cuisines (notably Chinese, Korean, European, and Japanese). Though neither meat nor vegetable, mushrooms are known as the "meat" of the vegetable world.^[21]

Most mushrooms sold in supermarkets have been commercially grown on mushroom farms. The most popular of these, *Agaricus bisporus*, is considered safe for most people to eat because it is grown in controlled, sterilized environments. Several varieties of *A. bisporus* are grown commercially, including whites, crimini, and portobello. Other cultivated species available at many grocers include *Hericium erinaceus*, shiitake, maitake (hen-of-the-woods), *Pleurotus*, and enoki. In recent years, increasing affluence in developing countries has led to a considerable growth in interest in mushroom cultivation, which is now seen as a potentially important economic activity for small farmers.^[22]

China is a major edible mushroom producer.^[23] The country produces about half of all cultivated mushrooms, and around 2.7 kilograms (6.0 lb) of mushrooms are consumed per person per year by over a billion people.^[24] In 2014, Poland was the world's largest mushroom exporter, reporting an estimated 194,000 tonnes (191,000 long tons; 214,000 short tons) annually.^[25]

Separating edible from poisonous species requires meticulous attention to detail; there is no single trait by which all toxic mushrooms can be identified, nor one by which all edible mushrooms can be identified. People who collect mushrooms for consumption are known as mycophagists,^[26] and the act of collecting them for such is known as mushroom hunting, or simply "mushrooming". Even edible mushrooms may produce allergic reactions in susceptible individuals, from a mild asthma-like response to severe anaphylactic shock.^{[27][28]} Even the cultivated *A. bisporus* contains small amounts of hydrazines, the most abundant of which is agaritine (a mycotoxin and carcinogen).^[29] However, the hydrazines are destroyed by moderate heat when cooking.^[30]

A number of species of mushrooms are poisonous; although some resemble certain edible species, consuming them could be fatal. Eating mushrooms gathered in the wild is risky and should only be undertaken by individuals knowledgeable in mushroom identification. Common best practice is for wild mushroom pickers to focus on collecting a small number of visually distinctive, edible mushroom species that cannot be easily confused with poisonous varieties.

Toxic mushrooms

Main article: [Mushroom poisoning](#)



Young *Amanitaphalloides*, "death cap" mushrooms

Many mushroom species produce secondary metabolites that can be toxic, mind-altering, antibiotic, antiviral, or bioluminescent. Although there are only a small number of deadly species, several others can cause particularly severe and unpleasant symptoms. Toxicity likely plays a role in protecting the function of the basidiocarp: the mycelium has expended considerable energy and protoplasmic material to develop a structure to efficiently

distribute its spores. One defense against consumption and premature destruction is the evolution of chemicals that render the mushroom inedible, either causing the consumer to vomit the meal (see emetics), or to learn to avoid consumption altogether. In addition, due to the propensity of mushrooms to absorb heavy metals, including those that are radioactive, European mushrooms may, as late as 2008, include toxicity from the 1986 Chernobyl disaster and continue to be studied.^{[31][32]}

Psychoactive mushrooms



Psilocybe zapotecorum, a hallucinogenic mushroom

Mushrooms with psychoactive properties have long played a role in various native medicine traditions in cultures all around the world. They have been used as sacrament in rituals aimed at mental and physical healing, and to facilitate visionary states. One such ritual is the *velada* ceremony. A practitioner of traditional mushroom use is the *shaman* or *curandera* (priest-healer).^[33]

Psilocybin mushrooms possess psychedelic properties. Commonly known as "magic mushrooms" or "shrooms", they are openly available in smart shops in many parts of the world, or on the black market in those countries that have outlawed their sale. Psilocybin mushrooms have been reported as facilitating profound and life-changing insights often described as mystical experiences. Recent scientific work has supported these claims, as well as the long-lasting effects of such induced spiritual experiences.^[34]



There are over 100 psychoactive mushroom species of genus *Psilocybe* native to regions all around the world.^[35]

Psilocybin, a naturally occurring chemical in certain psychedelic mushrooms such as *Psilocybe cubensis*, is being studied for its ability to help people suffering from psychological disorders, such as obsessive-compulsive disorder. Minute amounts have been reported to stop cluster and migraine headaches.^[36] A double-blind study, done by the Johns Hopkins Hospital, showed that psychedelic mushrooms could provide people an experience with substantial personal meaning and spiritual significance. In the study, one-third of the subjects reported ingestion of psychedelic mushrooms was the single most spiritually significant event of their lives. Over two-thirds reported it among their five most meaningful and spiritually significant events. On the other hand, one-third of the subjects

reported extreme anxiety. However, the anxiety went away after a short period of time.^[37] Psilocybin mushrooms have also shown to be successful in treating addiction, specifically with alcohol and cigarettes.^[38]

A few species in the genus *Amanita*, most recognizably *A. muscaria*, but also *A. pantherina*, among others, contain the psychoactive compound muscimol. The muscimol-containing chemotaxonomic group of *Amanita*s contains no amatoxins or phallotoxins, and as such are not hepatotoxic, though if not properly cured will be non-lethally neurotoxic due to the presence of ibotenic acid. The *Amanita* intoxication is similar to Z-drugs in that it includes CNS depressant and sedative-hypnotic effects, but also dissociation and delirium in high doses.

Mushroom Poisoning and treatments

Eating poisonous mushrooms may cause different types of reactions which can broadly be classified as follows :

1. **Gastric disorder:** The poison causes serious gastric disturbance, it chiefly acts by exciting and then paralyzing the central nervous system as by *Amanita muscaria* or poison containing irritant which cause gastric enteritis by direct action on the mucous membrane of the digestive system. e.g *Gyromitra esculenta*.
2. **Nervous disorder:** It causes degeneration of cells, especially of the nervous system and granular parenchymatous tissues like liver as in case of *Amanita phalloides*.
3. **Muscular disorder:** There may be exciting of the muscular system, especially the smooth muscular fibres as in the uterus, vessels etc.
4. **Haemolytic disorder:** There can be destruction of blood or haemolysis as in case of *Amanita rubescens*

Treatments :

- All the collectors of wild mushrooms should be careful about mushroom poisoning and have some knowledge of the first-aid remedies in case of mushroom poisoning and then the patient should immediately be taken to a doctor.
- The patient should be made to cover his body with a blanket, lie down calmly and give the first-aid treatment till the arrival of the doctor.
- **Removal of poison from the stomach:** The patient may be made to vomit by putting his fingers inside the mouth or throat or by giving warm water with one tablespoonful of mustard seeds or opomorphine. The stomach should be completely washed by means of a stomach tube. One can also give some sedatives like warm water, 4--5 tablespoonful of warm milk, two tablespoonful of olive oil beaten with the yolk of an egg etc.
- **Elimination of the toxin:** The ingested poison in the stomach can be removed by putting charcoal powder in the stomach and if it has already been absorbed in blood then subcutaneous injections of atropine or other antidotes can help in removing the effect of poisoning.

Mushrooms Cultivation: procedure for mushrooms cultivation!

Direct utilization of fungi as food:

Many Agaricales and Helvellales are directly used as food. There is a non-poisonous edible toadstool, i.e., *Coprinus* sp. found in lawns in the rainy season. *Agaricus campestris* is an edible mushroom and cultivated for its fruitifications. The fruiting bodies are quite fleshy and eaten directly as vegetable or with rice as 'pulao'.

KAHE



Image Courtesy: upload.wikimedia.org/wikipedia/commons/4/41/Mushroom-IMG_3304.JPG

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Prepared by Mr. R. Dineshkumar, Assistant Professor, Dept. of Microbiology, KAHE

These mushrooms are being successfully cultivated in South India. *Morchella esculenta* is another important edible fungus. It is found in Kashmir and Punjab plains. Its local name is 'guchi' and sold very costly. *Torulopsis utilis*, is used for the large-scale production of yeast for food purposes. *Saccharomyces cerevisiae* is used in breadmaking industry.

Of the many mushrooms that can be cultivated, only three kinds namely button mushroom (*Agaricus bisporus*), straw mushroom (*Volvariella volvacea*) and oyster mushroom (*Pleurotus sajorajju*) are suitable for growing in India where suitable environmental conditions exist. The following account deals with cultivation of Button Mushroom (*Agaricus bisporus*).

Growing Season:

Agaricus bisporus being a temperate mushroom grows best during winter throughout the plains of North India. It can however, be grown throughout the year in hills. The most suitable temperature for the spread of mycelium is 24-26°C. Temperature ranging from 16-18°C is essential for the formation of fruit bodies. High temperatures are harmful but the lower temperature retards the development of both mushroom mycelium and fruit bodies.

Mushroom House:

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The mushroom house can be any available room, shed, basement, garage, etc. The growing houses should be well ventilated and not stuffy.

Compost:

The cultivated mushroom is grown on special compost. Two types of composts, natural and synthetic are used for growing this mushroom.

Composting Yard:

The compost should be prepared on well cleaned concrete or pucca floor, which should be on a higher level so that the run-off water does not collect near the heap. Composting is usually done in the open, but it has to be protected from rain by covering it with polythene sheet. It can also be done in a shed with open sides or a larger room to shelter it from rain.

Synthetic Compost:

The following ingredients are required:

Wheat straw (chopped 8-20cm. long) – 250kg.

Wheat/Rice bran – 20kg.

Ammonium sulphate/calcium ammonium nitrate–3kg.

Urea–3kg.

Gypsum–20kg.

ADVERTISEMENTS:

This will make compost sufficient for 15-16 trays of size 1m×1/2m×15cm.

Ten kg molasses and/or 60 kg chicken manure can also be used if available.

Procedure for Long Method:

Mixing the materials and making pile: Day-0.

The straw is uniformly spread over the composting yard in a thin layer and wetted (thoroughly by sprinkling water). All ingredients such as wheat bran, fertilizers, etc., except gypsum are mixed thoroughly in the wetted straw, which is finally heaped into a pile about one metre high, one metre wide and as long as to hold the entire quantity of straw mixture. The pile can be made with hand or stack mould. The straw should be firmly but not compactly compressed into the mould.

It is essential to open the entire pile and remake it a number of times according to the following schedule:

1st turning–4th day

2nd turning–8th day

3rd turning–

12th day, add 10kg. gypsum.

4th turning final turning–16th day, add 10kg. gypsum.

Final turning–20th day, spray 10ml Malathion in 5litre water (any other available pesticide like, DDT, BHC, Lindane can also be used).

At each turning water should be sprinkled to make up the loss of water due to evaporation. If it is desired to add molasses, then 10kg molasses diluted 20 times with water should be poured over the straw mixture during the first turning. Sixty kg chicken manure if available can also be added at the time of start of pile, i.e., 0-day to improve the quality of the compost. Natural Compost

It is prepared from horse dung which must be freshly collected and not have been exposed to rain. It should not contain a mixture of dung of other animals. Chopped wheat straw (1/3, or more of weight of horse dung) is mixed with the horse dung to which 3 kg urea and/or 100-110 kg poultry manure per tonne is usually added.

Horse dung and straw mixture is uniformly spread over the composting yard and sufficient water is sprinkled over it so that the straw becomes sufficiently wet so that it takes no more water. The manure is then heaped in a pile as for synthetic compost.

After 3 days when the manure in the heap gets heated up due to fermentation and gives off an odour of ammonia, it is opened up and repeated 3 or 4 times after an interval of 3-4 days. Twenty five kg gypsum per tonne is added in two instalments at 3rd and 4th turning. At the final turning 20 ml Malathion diluted in 10 litres of water is sprayed into the manure.

Filling:

The compost when ready for filling and spawning has a dark brown colour and no trace of ammonia. There is no unpleasant odour but it smells like fresh hay. The pH is neutral or near neutral. The compost should not be too dry or too wet at the time of filling in the tray which can be determined by the palm test. For this purpose a small quantity of compost is taken into the hand and pressed lightly, if a few drops of water ooze from the fingers then it is of right consistency.

If relatively dry then the water should be added by sprinkling. If too wet, the excess water may be allowed to evaporate. The prepared compost is now filled in trays, which may be of any convenient size but its depth should be 15-18 cm. A standard size of tray 1 m x 1/2 m x 15 cm. The tray should be made of soft wood and provided with the pegs at the four corners so that they can be stacked one over the other leaving sufficient space between the two trays for various operations. The trays are completely filled with the compost, lightly compressed and the surface levelled.

Spawning:

Spawning means sowing the beds with the mycelium (spawn) of the mushroom. Spawn can be had from mushroom Laboratory College of Agriculture, Chamba, Solan at a nominal cost. Small quantity of spawn is also available from the Division of Mycology, Indian Agricultural Research Institute, New Delhi.

The grains of spawn are scattered on the surface of the tray bed which is covered with a thin layer of compost. Spawning can also be done by mixing the spawn with compost before filling it in trays. Five hundred gram spawn (two 1/2 lit bottles/poly packs) is sufficient for five trays of standard size. After spawning the compost surface is covered with old newspaper sheet, which is wetted by sprinkling water to provide the humidity but not water is directly added to the compost during spawning.

The trays after spawning are stacked vertically one over the other in 4-5 tiers. One metre clear space may be left in between the top tray and ceiling. There should be about 15-20 cm space between the two trays.

The room should be maintained at 25°C or near about. The humidity should be built up by frequently watering the floor and walls. The room may be kept closed as no fresh air is needed during the spawn run. White cottony mycelium spreads and permeates through the compost.

Eventually the compost surface gets covered with the mycelium. It takes 12-15 days for complete spawn run. Low temperature prolongs the spread of the mycelium.

Casing:

After the spawn run is complete as is evident by white mouldy growth, the surface of the compost is covered with 3 cm layer of casing soil. A suitable casing soil can be prepared by mixing equal part of well rotted cow dung (finely crushed and coarsely sieved) and garden soil. The casing material should possess high water holding capacity, good pore space and pH should not be lower than 7.4. The casing material is sterilized to kill insects, nematodes and molds. Sterilization can be accomplished either by steaming or by treating with formalin solution. For one cubic metre of casing soil, half litre of formalin diluted with 10 litre of water is sufficient.

The casing soil is spread over a plastic sheet and treated with formalin by sprinkling. The treated soil is piled up in a heap and covered with another plastic sheet for 48 hours. The soil is turned frequently for about a week to remove all traces of formalin which can be tested by smelling it. After casing, the temperature of the room is maintained for further three days, after which it must be lowered to below 18°C. At this stage lot of fresh air is needed and therefore, the growing room should be ventilated by opening windows, etc.

Cropping and Harvesting:

The first flush of the pinheads becomes visible 15-20 days after casing or say about 35-40 days after spawning. Small white buttons develop 5-6 days after pinhead stage. The right stage of harvest is when the cap is still tight over the short stem. In case the buttons are allowed to mature, the membrane below the cap will rupture and the cap will open up in umbrella-like shape.

Such mushrooms are considered to be inferior. Harvesting is done by holding the cap with forefingers slightly pressed against the soil. The soil particles and mycelial threads clinging to the base of the stalk are chopped off. Mushroom can also be harvested by cutting off with a sharp knife at soil level.

Yield:

The average yield of 3-4 kg per tray is considered normal. However, if compost is carefully prepared, spawn reliable and favourable temperature prevailing during the growing period, then a yield of 5-6 kg per tray is possible. Partial or complete failure may also happen due to negligence.

Storage:

The mushrooms are best consumed fresh. Storage in refrigerator for a few days is possible. The mushrooms should be placed between moist paper towel for storing in a refrigerator.

Classification of Mushrooms

- Mushroom is a fleshy fruiting body of some fungi arising from a group of mycelium buried in substrate. Most of the mushrooms belong to the Sub-Division: Basidiomycotina and a few belong to Ascomycotina of Kingdom-Fungi.
- It is reported that there are about 50,000 known species of fungi and about 10,000 are considered as edible ones. Of which, about one hundred and eighty mushrooms can be tried for artificial cultivation and seventy are widely accepted as food. The cultivation techniques were perfected for about twenty mushrooms and about dozen of them have been recommended for commercial cultivation. However, only six mushrooms are widely preferred for large-scale cultivation. They are:
 1. Paddy straw mushroom - *Volvariella* spp.
 2. Oyster mushroom - *Pleurotus* spp.
 3. Button mushroom - *Agaricus* spp.
 4. Milky mushroom - *Calocybe* spp.
 5. Shiitake mushroom - *Lentinula* spp.
 6. Jew's ear mushroom - *Auricularia* spp.

MUSHROOM CULTIVATION - 17MBU601A Possible Questions – Unit I

1. What is a mushroom?
2. Write the general morphology of button mushroom?
3. Write about mother spawn?
4. What is mushroom Nutraceuticals?
5. How to preserve the mushroom?
6. Write about the variations in mushroom morphology.
7. Explain Ainsworth et al classification of mushroom.
8. Explain the spore germination & life cycle of *Agaricus bisporus*.

SINO	unit-1	Option A	Option B	Option C	Option D	Answer
1	Mushroom consist of _____ portions	1	2	3	4	2
2	Two portions of Mushrooms are _____	Fruiting bodies and mycelium	mycelium and thread like structures	stalks and fruiting bodies	stalk and mycelium	Fruiting bodies and mycelium
3	Many mushrooms have _____	Fruiting bodies and mycelium	mycelium and thread like structures	stalks and fruiting bodies	caps and mycelium	stalks and caps
4	_____ supplies nutrients from substrate to fruiting bodies	Stalks	chlorophylls	caps	mycelium	mycelium
5	How many species of mushrooms are cultivated in india?	10	25	23	35	25
6	A ring is formed around the stipe is called as _____	annulus	veil	gills	pileus	annulus
7	The _____ covers the gills and extend from the margin of the cap to the stalk.	annulus	veil	gills	pileus	veil
8	Gills are also called as _____	annulus	veil	gills	lamellae	lamellae
9	In many varieties, fruiting bodies remain covered by a _____	annulus	veil	gills	pileus	veil
10	The stalk arises from a cuplike structure known as _____	volva	veil	gills	pileus	volva
11	The _____ and _____ are supported by the stipe(Stalk)	pileus and gills	annulus and lamellae	veil and cap	none of the above	pileus and gills
12	The genus _____ has a funnel shaped fruiting body with folds	Cantharellus	hymenium	Schizophyllum	volverella	Cantharellus
13	_____ is the example of stripe less mushroom.	Cantharellus	hymenium	Schizophyllum	volverella	Schizophyllum
14	Forming fruiting bodies entirely above the _____	Substratum	tuber	truffles	tuberaceae	Substratum

	surface of _____					
15	On the basis of the occurrence of fruiting bodies, mushrooms may be classified into two groups called _____	Epigenous	hypogenous	epigenous and hypogenous	pizza species	epigenous and hypogenous
16	Gilled fungi have been further divided into _____ categories.	3	1	5	8	5
17	Pore fungi is characterized by absence of _____	gills	tubes	tubes or pores	pores	gills
18	Pore fungi is characterized by presence of _____	gills	tubes	tubes or pores	pores	tubes or pores
19	Pore fungi is characterized by _____ gills.	absence	presence	absence or presence	none of the above	absence
20	Pore fungi is characterized by _____ Tubes or pores.	absence	presence	absence or presence	none of the above	absence
21	Pore fungi is characterized by _____ Tubes or pores	Very hard and irregular	Woody and irregular	Regular and irregular	Very hard and woody	Very hard and woody
22	Pores mating the context in an _____ and _____	Very hard and irregular	Woody and irregular	Regular and irregular	Irregular and uneven line	Irregular and uneven line
23	In <i>Boletaceous</i> whose many species can be distinguished among them only by _____ characteristics	Microscopic	naked eye	macroscopic	none of the above	microscopic
24	In <i>Clavaria cinerea</i> fruit bodies are _____ in colour.	Pale violet	pale yellow	ashgrey	red	ashgrey
25	In <i>Clavaria flava</i> flesh of pileus and its branches turning _____ brushed	Pale violet	pale yellow	ashgrey	red	red
26	In <i>Clavaria amethystina</i> fruit bodies are _____ in colour	Pale violet	pale yellow	ashgrey	red	Pale violet
27	In <i>Clavaria aurea</i> flesh not turning _____ when brushed	Pale violet	pale yellow	ashgrey	red	red

28	In <i>Clavaria pistillaris</i> fruit bodies _____ in colour.	Pale violet	pale yellow	ashgrey	tan	tan
29	The most widely cultivated mushroom is	<i>Agaricus bisporus</i>	<i>Volvariella volvacea</i>	<i>Pleurotus ostreatus</i>	Both A and B	<i>Agaricus bisporus</i>
30	<i>Pleurotus ostreatus</i> commonly called as	Oyster mushroom	Button mushroom	Paddy straw mushroom	Magic mushroom	oyster mushroom
31	<i>Morchella</i> species are _____ mushrooms.	Ascomycetous	Myceteous	gastromycetous	none of the above	Ascomycetous
32	<i>Morchella</i> species are also called as _____	morels	corals	sponge	none of the above	morels
33	<i>Hydnum caputursi</i> fries mushroom is also known as _____	Bear's head	Hydunum	both a and b	edible	both a and b
34	Poison mushroom is classified by_____	8	6	5	2	5
35	Muscarine poisoning is known to be caused by the eating of _____	<i>Amanita muscaria</i>	<i>Amanita pantherina</i>	<i>Amanita vaginata</i>	none of the above	<i>Both A and B</i>
36	Haemolytic poisoning is known to be caused by the eating of _____	<i>Amanita muscaria</i>	<i>Amanita rubescens</i>	<i>Amanita pantherina</i>	<i>Amanita vaginata</i>	<i>Amanita vaginata</i>
37	Psychotropic poisonous mushroom affects	UTI	heart	kidney	nervous system	nervous system
38	Coprine poisonous mushroom affects	UTI	heart	kidney	nervous system	nervous system
39	What is the other name of Mushroom?	Funaria	agaricus	dryopteris	fern	<i>Agaricus</i>
40	Mushroom is:	Saprophytic fungus	Autotrophic Algae	Heterotrophic fungus	none of the above	Saprophytic fungus
41	The scientific name of paddy straw mushroom is	<i>volvariella volvacea</i>	<i>Calocybe indica</i>	<i>Agaricus bisporus</i>	<i>Pleurotus sojarcaju</i>	<i>volvariella volvacea</i>
42	The nutritive value of paddy straw mushroom is _____%protein.	90	40	6	35	
43	<i>Agaricus bisporus</i> can grow in _____	tropical region	temperate region	any type	desert	temperate region

44	Mushroom is a	Saprophytic fungus	halophyte	parasite	autotroph	Saprophytic fungus
45	Which of the following statement is true?	normal soil is suitable	in any types of soil	grown in particular types of soil in temperate region	cannot be cultivated in south india	grown in particular types of soil in temperate region

K A H E

Unit II

Biology of Mushrooms: Vegetative characters, general morphology, spore germination and life cycle of button mushroom (*Agaricus bisporus*), milk mushroom (*Calocybe indica*), oyster mushroom (*Pleurotus sajorajju*) and paddy straw mushroom (*Volvariella volvacea*).

OYSTER MUSHROOM CULTIVATION PROCESS

MATERIAL REQUIREMENT

Substrate:

A large number of agriculture, forest and agro-industrial byproducts are useful for growing oyster mushroom. Substrates should be fresh, dry and free from mould infestation. Oyster mushroom can utilize a large number of agro-wastes including straw of wheat, paddy and ragi, stalks and leaves of maize, jowar, bajra, and cotton, sugarcane bagasse, jute and cotton waste, peanut shells, dried grasses, sunflower stalks, used tea leaf and discarded waste paper. It can also be cultivated using industrial wastes like paper mill sludge, coffee by-products, tobacco waste etc. About 1.5-2.0 kg of good substrate will be required per bag of 80 cm x 40 cm size.

Mushroom Spawn

Three to four week old non-contaminated spawn @ 10% of dry weight of the substrate is required for the purpose. Spawn of good quality should be collected from a reliable source. Further, the species/variety should be chosen basing on the temperature and relative humidity of the cropping season of the locality. Just prior to use the 200 gm spawn is extracted from the bottle with hooked iron rod and divided into four parts.

Substrate Supplementation

Some of the common supplements are wheat bran, rice bran, soybean cake, groundnut cake, maize meal, horse gram powder, cotton seed meal etc. wheat bran and rice bran should be used at the rate of 10%, while others should be tried @ 3-6% on dry weight basis of the substrate. Supplements after pasteurization are thoroughly mixed with straw while spawning. Addition of supplements increases the substrate temperature and hence, it is risky during the work period to supplement the substrate.

Paddy straw mushroom-Bed preparation and cropping

The cultivation of paddy straw mushroom can be done in a thatched house and also under the shade of a tree. Fresh, disease free paddy straw is the ideal substrate. Ten-fifteen kg paddy straw is necessary for preparing one

bed. In recent years, it is cultivated inside plastic film houses to maintain the temperature of around 25-35°C and relative humidity of 75-80%.

Paddy Straw bundle method

Procedure

- Prepare a raised platform of about 1m in length and 0.75m in breadth with a deal wood flanks and keep it over a support by arranging bricks on all four corners.
- The paddy straw is bundled into two weigh about 1kg each.
- Soak the straw bundles in water for 12-18hr.
- Ten bundles are taken out and drain the excess water
- Place the bundles over the platform with their butt end on one side.
- Build the second layer by placing the butt end towards the other direction. (These 8 bundles make one layer of bed)
- Place the small quantity of spawn 8-12cm inside the margin at an interval of 10-15cm all along the periphery.
- Apply a spoonful of coarsely powdered dhall powder before placing spawn.
- Place the straw bundles at right angle to the previous layer in criss-cross fashion to make the third layer.
- Place the straw bundles with opposite butt end to make fourth layer.
- Spawn this layer as stated above.
- Place another layer of straw bundles over this and do not apply spawn.
- Pressure the bed to make it as compact as possible and cover it with a transparent polythene sheet.
- Keep the beds undisturbed for a few days.

Note: Usually the bed will have the necessary humidity, if the straw bundles are properly soaked. If moisture is found to be less, it may be watered using a rose can. On the other hand, if the moisture is found to be excess, polythene cover is to be partly to regulate the moisture. The success of cultivation depends upon the temperature and moisture in the bed. The optimum temperature of 30-35°C is necessary for developing buttons.

The mushrooms start appearing from all sides in 6-10 days as tiny buttons, which can be harvested in another 4-5 days. The harvesting is to be done at the button stage itself, since the opened sporocarp will be more fibrous. Usually, 1-2kg of mushroom can be harvested from 10kg substrate.

Paddy straw twist method

Instead of bundled straw, twisted paddy straw can also be used for cultivation

Procedure

- Make the straw into twists of about 5-8m long and 5-10cm diameter.
- Immerse the twists in water for 12hr.
- Take out the straw and drain the excess water.
- Place the twists lengthwise over a platform in a zigzag manner.
- Place a second over this in an opposite direction. (This forms the first layer of the bed)
- Sprinkle the coarsely powdered dhall and place small bits of spawn all along the periphery as above.
- Build another layer as described above and spawn the layer.
- Build up 4-5 layers and spawn as usual.
- Compact the bed by pressing and cover it with a polythene sheet.

Button Mushroom

Taxonomy

The common mushroom has a complicated taxonomic history. It was first described by English botanist Mordecai Cubitt Cooke in his 1871 *Handbook of British Fungi*, as a variety (var. *hortensis*) of *Agaricus campestris*.^{[4][5]} Danish mycologist Jakob Emanuel Lange later reviewed a cultivar specimen, and dubbed it *Psalliotahortensis* var. *bisporus* in 1926.^[6] In 1938, it was promoted to species status and renamed *Psalliotabispora*.^[7] Emil Imbach (1897–1970) imparted the current scientific name of the species, *Agaricus bisporus*, after the genus *Psalliota* was renamed to *Agaricus* in 1946.^[3] The specific epithet *bisporus* distinguishes the two-spored basidia from four-spored varieties.

Description

The pileus or cap of the original wild species is a pale grey-brown in color, with broad, flat scales on a paler background and fading toward the margins. It is first hemispherical in shape before flattening out with maturity, and 5–10 centimetres (2–4 inches) in diameter. The narrow, crowded gills are free and initially pink, then red-brown and finally a dark brown with a whitish edge from the cheilocystidia. The cylindrical stipe is up to 6 cm (2½ in) tall by 1–2 cm wide and bears a thick and narrow ring, which may be streaked on the upper side. The firm flesh is white, although it stains a pale pinkish-red on bruising.^{[8][9]} The spore print is dark brown. The spores are oval to round and measure approximately 4.5–5.5 µm × 5–7.5 µm, and the basidia usually two-spored, although two-tetrasporic varieties have been described from the Mojave Desert and the Mediterranean, with predominantly heterothallic and homothallic lifestyles, respectively.^{[10][11]}

This mushroom is commonly found worldwide in fields and grassy areas following grain, from late spring through to autumn, especially in association with manure. It is widely collected and eaten, even by those who would not normally experiment with mushroom hunting.^[9]

Similar species

The common mushroom could be confused with young specimens of the deadly poisonous destroying angel (*Amanita* sp.), but the latter may be distinguished by their volva or cup at the base of the mushroom and pure white gills (as opposed to pinkish or brown of *A. bisporus*). Thus it is always important to clear away debris and examine the base of such similar mushrooms, as well as cutting open young specimens to check the gills. Furthermore, the destroying angel grows in mossy woods and lives symbiotically with spruce.

A more common and less dangerous mistake is to confuse *A. bisporus* with *Agaricus xanthodermus*, an inedible mushroom found worldwide in grassy areas. *A. xanthodermus* has an odor reminiscent of phenol; its flesh turns yellow when bruised. This fungus causes nausea and vomiting in some people.

The poisonous European species *Entoloma sinuatum* has a passing resemblance as well, but has yellowish gills, turning pink, and it lacks a ring.

Cultivation history



A. bisporus being cultivated

The earliest scientific description of the commercial cultivation of *A. bisporus* was made by French botanist Joseph Pitton de Tournefort in 1707.^[12] French agriculturist [Olivier de Serres](#) noted that transplanting mushroom mycelia would lead to the propagation of more mushrooms.

Originally, cultivation was unreliable as mushroom growers would watch for good flushes of mushrooms in fields before digging up the mycelium and replanting them in beds of composted manure or inoculating 'bricks' of compressed litter, loam, and manure. Spawn collected this way contained pathogens and crops commonly would be infected or not grow at all.^[13] In 1893, sterilized, or pure culture, spawn was discovered and produced by the Pasteur Institute in Paris, for cultivation on composted horse manure.^[14]

Today's commercial variety of the common mushroom originally was a light brown color. In 1926, a Pennsylvania mushroom farmer found a clump of common mushrooms with white caps in his mushroom bed. As with the reception of white bread, it was seen as a more attractive food item and became very popular.^[15] Similar to the commercial development history of the [navel orange](#) and [Red Delicious apple](#), cultures were grown from the mutant individuals, and most of the cream-colored store mushrooms marketed today are products of this 1926 chance natural mutation.

A. bisporus is now cultivated in at least seventy countries throughout the world.^[3] Global production in the early 1990s was reported to be more than 1.5 million short tons (1.4 billion kilograms), worth more than US\$2 billion.^[16]

Nutritional profile

Agaricus bisporus, white raw

Nutritional value per 100g (3.5oz)

Energy 93kJ (22kcal)

Carbohydrates 3.26g

Sugars 1.98g

Dietary fiber 1g

Fat 0.34g

Protein 3.09g

Vitamins Quantity %DV[†]

Thiamine(B1)	7%	0.081mg
Riboflavin(B2)	34%	0.402mg
Niacin(B3)	24%	3.607mg
Pantothenic acid(B5)	30%	1.497mg
VitaminB6	8%	0.104mg
Folate(B9)	4%	17µg
VitaminB12	2%	0.04µg
VitaminC	3%	2.1mg
VitaminD	1%	0.2µg

Minerals Quantity %DV†

Iron	4%	0.5mg
Magnesium	3%	9mg
Phosphorus	12%	86mg
Potassium	7%	318mg
Sodium	0%	3mg
Zinc	5%	0.52mg

Other constituents Quantity

Water	92.45g
-------	--------

Link to USDA Database entry

•Units
•µg= micrograms •mg= milligrams
•IU= International units

†Percentages are roughly approximated using

US recommendations for adults.

Source: USDA Nutrient Database

In a 100-gram serving, raw white mushrooms provide 93 kilojoules (22 kilocalories) of food energy and are an excellent source (>19% of the Daily Value, DV) of the B vitamins, riboflavin, niacin, and pantothenic acid (table). Fresh mushrooms are also a good source (10–19% DV) of the dietary mineral phosphorus (table).

While fresh *A. bisporus* only contains 0.2 micrograms (8 IU) of vitamin D as ergocalciferol (vitamin D₂), the ergocalciferol content increases substantially after exposure to UV light.^{[17][18]}

Polythene Bag

Transparent polythene tube of 125-150 gauge with a dimension of 80 cm x 40 cm is suitable for oyster cultivation. Bags of 60 cm x 40 cm may also be used for the purpose. The bags can be reused for the second crop after proper cleaning.

CULTIVATION PROCEDURE

Substrate Processing

Freshly procured good quality substrate is chopped to 4-5 cm size by a chaff cutter and steeped in a chemical solution of carbendazim 50% WP (75 ppm) and formaldehyde (500 ppm) for a period of 6 hours. Then the straw is taken out and excess water is to be drained. Ninety liters of water mixed with 7.5 g carbendazim 50% WP and 125 ml formaldehyde (37-40%) will be appropriate for the purpose. However, steam and hot water treatment methods are preferred as there are reports of phytotoxicity associated with chemical treatment. Here, the pre-wetted substrate after chopping is soaked in hot water (65-70°C) for one hour. In case of steam pasteurization, the pre-wetted straw is steam pasteurized at 60-70°C for one hour, cooled at room temperature and then seeded with spawn. Through pasteurization, the competitor moulds are either killed or their growth is suppressed for 25-40 days after spawning.

The substrate is dried in shade for few hours in order to maintain the moisture level of 55-60%. The substrate necessary for raising one bag may be divided into four lots after drying.

Raising of bag

One end of the polythene tube is tied with a rubber band and the moistened substrate is put inside to a height of 15 cm. Substrate is then gently pressed and one part each of spawn (50g) and supplement (50g) spread at the periphery

close to polythene (See the Video). Likewise, four such layers are made

and the bag is closed at the upper end after pressing the substrate. For a bag out of 2kg of dry straw, 200g each spawn and supplement will be used. 15-

20 small holes (0.5cm diameter) should be made on all sides to facilitate gas exchange. Instead of layers spawning, mixed spawning may also be followed where the required quantity of spawn is mixed with the prepared substrate (soaked straw) and incorporated into the bag. The bags are then incubated in a well ventilated room at 25

C. During the mycelial growth bags should not be opened.

After Care

Once the mycelium has fully colonized the substrate forming a thick mycelial mat, it is ready for fruiting. Contaminated bags with mould may be discarded while bags with patchy mycelial growth may be left for few more days for completion of the mycelial growth. These bags are opened after 15-16 days. But in case of *P. oous* and *P.*

djamorevar. are opened after 12 days as fruiting comes out within these. The bundles after opening are arranged on shelves at a distance of 20cm between each bag in the tier or hanged with plastic rope. Appropriate temperature (20-30°C), humidity (70-80%) and light (200lux) with good ventilation should be maintained in the cropping room.

Bags are watered twice daily depending upon the weather condition.

Harvesting

Primordia (small eggs) appears within 4-5 days of opening the bag that came to the harvestable stage 3-4 days later. The mushroom should be harvested when the cap begins to fold inwards. Picking is done by twisting the mushroom gently without disturbing the surrounding fruit bodies. Crop should not be watered before harvesting.

The second crop appears after 7-10 days. Hence within 45 days crop period, 3-4 crops are expected. With exception, under suitable growing condition, a biological efficiency of 100% is achieved in commercial farms.

Mushrooms Cultivation: procedure for mushrooms cultivation!

Direct utilization of fungus as food:

Many Agaricales and Helvellales are directly used as food. There is a non-poisonous edible toadstool, i.e., *Coprinus* sp. found in lawns in the rainy season. *Agaricus campestris* is an edible mushroom and cultivated for its fructifications. The fruiting bodies are quite fleshy and eaten directly as vegetable or with rice as 'pulao'.

These mushrooms are being successfully cultivated in South India. *Morchella esculenta* is another important edible fungus. It is found in Kashmir and Punjab plains. Its local name is 'guchi' and sold very costly. *Torulopsis utilis*, is used for the large-scale production of yeast for food purposes. *Saccharomyces cerevisiae* is used in bread making industry.

Of the many mushrooms that can be cultivated, only three kinds namely button mushroom (*Agaricus bisporus*), straw mushroom (*Volvariella volvacea*) and oyster mushroom (*Pleurotus sajorajju*) are suitable for growing in India where suitable environmental conditions exist. The following account deals with cultivation of Button Mushroom (*Agaricus bisporus*).

Growing Season:

Agaricus bisporus being a temperate mushroom grows best during winter throughout the plains of North India. It can however, be grown throughout the year in hills. The most suitable temperature for the spread of mycelium is 24-26°C. Temperature ranging from 16-18°C is essential for the formation of fruit bodies. High temperatures are harmful but the low temperature retards the development of both mushroom mycelium and fruit bodies.

Mushroom House:

The mushroom house can be any available room, shed, basement, garage, etc. The growing houses should be well ventilated and not stuffy.

Compost:

The cultivated mushroom is grown on special compost. Two types of composts, natural and synthetic are used for growing this mushroom.

Composting Yard:

The compost should be prepared on well cleaned concrete or pucca floor, which should be on a higher level so that the runoff water does not collect near the heap. Composting is usually done in the open, but it has to be protected

from rain by covering it with polythene sheet. It can also be done in a shed with open sides or a larger room to shelter it from rain.

Synthetic Compost:

The following ingredients are required:

Wheat straw (chopped 8-20 cm. long) – 250 kg.

Wheat/Rice bran – 20 kg.

Ammonium sulphate/calcium ammonium nitrate – 3 kg.

Urea – 3 kg.

Gypsum – 20 kg.

The casing soil is spread over a plastic sheet and treated with formalin by sprinkling. The treated soil is piled up in a heap and covered with another plastic sheet for 48 hours. The soil is turned frequently for about a week to remove all traces of formalin which can be tested by smelling it. After casing, the temperature of the room is maintained for further three days, after which it must be lowered to below 18°C. At this stage lot of fresh air is needed and therefore, the growing room should be ventilated by opening windows, etc.

Cropping and Harvesting:

The first flush of the pinheads becomes visible 15-20 days after casing or say about 35-40 days after spawning. Small white buttons develop 5-6 days after pinhead stage. The right stage of harvest is when the cap is still tight over the short stem. In case the buttons are allowed to mature, the membrane below the cap will rupture and the cap will open up in umbrella-like shape.

Such mushrooms are considered to be inferior. Harvesting is done by holding the cap with forefingers slightly pressed against the soil. The soil particles and mycelial threads clinging to the base of the stalk are chopped off. Mushroom can also be harvested by cutting off with a sharp knife at soil level.

Yield:

The average yield of 3-4 kg per tray is considered normal. However, if compost is carefully prepared, spawn reliable and favourable temperature prevailing during the growing period, then a yield of 5-6 kg per tray is possible. Partial or complete failure may also happen due to negligence.

Storage:

The mushrooms are best consumed fresh. Storage in refrigerator for a few days is possible. The mushrooms should be placed between moist paper towel for storing in a refrigerator.

Milk mushroom

White milky mushroom (*Calocybe indica*)

Milk mushroom (*Calocybe indica*) can be grown on a wider range of substrates as in case of oyster mushroom. It can be grown on substrates containing lignin, cellulose and hemicelluloses. Substrates should be fresh and dry. Substrates exposed to rain or harvested premature (green color) are prone to various weed moulds which may result in failure of the crop. It can be grown on straw of paddy, wheat, ragi, maize/bajra/cotton stalks and leaves, sugarcane bagasse, cotton and jute wastes, dehulled maize cobs, tea/coffee waste etc., However cereal straw (paddy/wheat) easily available in abundance, is being widely used.

Straw is chopped in small pieces (2-4 cm size) and soaked in fresh water for 8-16 hours. This period can be reduced when pasteurization is to be done by steam. Main purpose of soaking is to saturate the substrate with water. It is easier to soak if straw is filled in gunny bag and dipped in water.



Pasteurization

The purpose of pasteurization is to kill harmful microbes. This can be achieved in two ways.

Hot water treatment

Water is boiled in a wide mouth container and chopped wet straw filled in gunny bags is submerged in hot water for 40 minutes at 80-90°C to achieve pasteurization. This is a very popular method particularly with small growers.

Steam pasteurization

Wet straw is filled inside an insulated room either in perforated shelves or in wooden trays. Steam is released under pressure from a boiler and temperature inside substrate is raised to 65°C and maintained for 5-6 hours. Air inside the room should be circulated to have uniform temperature in the substrate.

Sterilization

Substrate is filled in polypropylene bags (x45cm, holding 2-3kg wet substrate) and sterilized at 15 lbsi for 1 hour. Once pasteurization/sterilization is over, straw is shifted to spawning room for cooling, bag filling and spawning.



Spawning and spawn running

Spawning methods are similar to that mentioned in case of oyster mushroom. In case of sterilization of PP bags, autoclave either surface or through spawning should be done. Higher spawn dose 4-5% of wet substrate is used. After spawning, bags are shifted to spawn running room and kept in dark where temperature 25-35°C and relative humidity above 80% are maintained. It takes about 20 days when substrate is fully colonized and bags are ready for casing. Bags are shifted to cropping room for casing and

cropping.



Casing

Casing means covering the top surface of bags after spawn run is over, with pasteurized casing material in thickness of about 2-3 cm. Casing provides physical support, moisture and allows gas to escape from the substrate. Casing material (soil 75% + sand 25%) with pH adjusted to 7.8-7.9 with chalk powder is pasteurized in autoclave at 151 psi for one hour or chemically treated with formaldehyde solution (4%) about a week in advance of casing. Solutions should be enough to saturate the soil. It is covered with polythene sheet to avoid escape of chemical and at an interval of 2 days soil is turned so that at the time of casing soil is free from formalin fumes. Bag's top is made uniform by ruffling top surface of the substrate and sprayed with solution of carbendazim (0.1%) + formaldehyde (0.5%). Casing material is spread in uniform layer of 2-3 cm thickness and sprayed with solution of carbendazim and formaldehyde to saturation level. Temperature 30-35°C and R.H. 80-90% are maintained.

Cropping

It takes about 10 days for mycelium to reach on top of casing layer when fresh air is introduced while maintaining temperature and R.H. as above. Light should be provided in long time. The changes thus made in environment, result in the initiation of fruiting bodies within 3-5 days in the form of needleshape which mature in about a week. Mushrooms 7-8 cm diam. are harvested by twisting, cleaned and packed in perforated polythene/polypropylene bags for marketing. Mushrooms can also be wrapped in kiln film for longer storage.

MUSHROOM CULTIVATION-17MBU601A

POSSIBLE QUESTIONS

1. Write a detailed note on production of mother spawn & its multiplication.
 2. Explain the harvesting, packaging, storage and marketing of mushrooms.
 3. What are the different parts of a typical mushroom?
 4. What is milky mushroom?
 5. How to isolate & culture the spores for mushroom cultivation?
 6. Write about the anti-tumor effect of mushroom.
 7. Write the steps involved in post harvest technologies of mushroom cultivation.
 8. Explain the vegetative characters & life cycle of milky mushroom.
 9. Explain the vegetative characters & life cycle of paddy straw mushroom.
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SL.NO	unit-2	Option A	Option B	Option C	Option D	Answer
1	The presence of spore bearing layer known as	mycelium	hymenium	rhizomorph	sporangium	hymenium
2	The hyphae of secondary mycelium twist together to form wide hyphal cords called,	mycelium	hymenium	rhizomorph	sporangium	rhizomorph
3	Rhizomorphs are visible in which stage	red hyphal strands	white hyphal strands	green hyphal strands	none of the above	green hyphal strands
4	The hyphal Swelling or ovoid body called	button stage	mycelial aggregation	chymatous	sporophore	button stage
5	Which type of genera the young button is covered by a universal veil	Amanita	button mushroom agaricus	Paddy straw	milky mushroom	Amanita
6	the central part of gill which is made up of ____	mycelium, hyphea	trema,mycelial threads	hymenium, sub hymenium	trema, basidia	trema,mycelial threads
7	Basidia are _____ cells which are arranged parall	long club shaped	short club shaped	long chain shaped	short chain shaped	long club shaped
8	White button mushroom known as	agaricus bisporus	lentinus edodes	agaricus campestris	basidiospores	agaricus bisporus
9	Agaricus biaporus is grown on	cereal straw	animal manure whea	cereal straw, animal manure	none of the above	cereal straw, animal manure
10	Most extensively cultivated mushroom in the world is	agaricus bispours	milky mushroom	oyster mushroom	button mushroom	agaricus bispours
11	The cropping period of Lentinus edodes	3 to 5 years	4to 6	2 to 5	4 to 8	3 to 5 years

12	Oyster mushroom also known as	flammulina velutipes	volvariella volvacea	pleurotus sps	pleurotus cystidus	pleurotus sps
13	Fruiting structure can be produced by	single monosporous mycelium	monosporous mycelium	polysporous mycelium	disporous mycelium	single monosporous mycelium
14	Pezizales order contains approximately _____ species and ____ families.	102 species, 16 families	1125 species , 16 familie	16 species,225 families	1025 species, 16 families	1125 specices , 16 familie
15	_____ mushrooms contain self-fertile and self-sterile species	Edible mushrooms	Ascomycetes	discomycetes	basidiocarp	Edible mushrooms
16	which mushroom is grown in high tempertature and tropical regions	pleurotus species	Volvariella volvacea	flammulina velutipes	Agaricus bisporus	Volvariella volvacea
17	which mushroom is contain the high amount od pro- vitamins and vitamin D	pleurotus species	Volvariella volvacea	flammulina velutipes	Agaricus bisporus	flammulina velutipes
18	Truffle belong to	Ascomycetes	eumycota	basidiomycete	discomycetes	Ascomycetes
19	Black cup fungus is	truffles	plectania campylospora	morchellaceae	pithya vulgaris	plectania campylospora
20	Poisonous mushrooms may be identified based on	morphology	colour	Shape	having swollen stacks	having swollen stacks
21	which is the highly toxic mushroom	Amanita virosee	Gyromitra esculenta	A.phalloids	Agaricus	A.phalloids
22	which mushroom toxin is Water soluble	Gyromitra esculenta	amanita phalloides	A.muscaria	A.verna	Gyromitra esculenta
23	A.muscuria is toxic cause	mycetomuscarine	muscardine	Diarrhoea	Mycetomuscarine and muscardine	Mycetomuscarine and muscardine
24	what is the Elimination of	the toxin is mixed	it caues	the toxin	.the toxin is absorbed in	.the toxin is absorbed in

	toxin	in the blood	vomitting	disturbances respiratory track	blood and eliminated by subcutaneous injection of atropin	blood and eliminated by subcutaneous injection of atropin
25	Pleurotus mushroom is _____ dependant	water	temperature	Nutrient	light	light
26	Tetrapolar mating system, the mating competence is determined by incompatibility factors are	factor A and B	A and c	factor AB	none of the above	factor A and B
27	Winter mushroom known as	pleurotus species	Volvariella volvacea	flammulina velutipes	Agaricus bisporus	flammulina velutipes
28	Phallales looking like a puffball because they are incased in an outer skin known as	spores	Peridium	peridioles	spore masa	Peridium
29	Saffron milk cap is otherwise called	morchella species	boletus mellea	Russula specia	lactarius deliciosus	lactarius deliciosus
30	Consumption of Psilocybecubensis appear in	30 minutes	30-60 minute	. 20 minutes	within few minutes	30-60 minute
31	Amanita muscuria and A.pantherina mushrooms contain	protein	muscarine and ibotenic acid	norcaperatic acid	psilocybin	muscarine and ibotenic acid
32	How to differentiate edible mushroom and poisonous mushroom	shape	small	Colour	size	colour
33	Amanita and phalloidin toxin producing mushroom	Amanitaphalloides	A.muscuria	angel mushroom	Gyromitraesculenta	Amanitaphalloides
34	Which types of mushroom is has no cap, gills or stems	Hymenogastrales	Nidulariales	Lycoperdales	phallales	Nidulariales

35	Paddy straw is made into bundles having ----- diameter	2 to 4 diameter	3 diameter	0.80 to 1.00 diameter	0.5 to 1 diameter	0.80 to 1.00 diameter
36	Moisture contain in mushroom	85%	90%	69%	45%	90%
37	Mushroom lack of	protein	aminoacids	Chorophyll	toxin	chorophyll
38	Consumption of Psilocybecubensis appear in	30 minutes	30-60 minutes	20 minutes	within few minute	30-60 minutes
39	SCP contain ----% protein	55%	45%	75%	100%	45%
40	Which of the following is a good compost ?	wet compost	with offensive smell	with Ph 4	with pH 7 and no offensive smell of ammonia	with pH 7 and no offensive smell of ammonia
41	Mushroom cultivation was started first in	France	China	England	Italy	France
42	Spawn is :	mycelium of fungus	fruiting body of fungus	type of compost	thin layer of soil forming	mycelium of fungus
43	For oyster mushroom cultivation beds are prepared in	Plastic bags	trays	Pots	buckets	Plastic bags
44	The mushroom spawn preparation needs addition of calcium carbonate because it	maintain Ph	absorbs excess water	helps in grains separate	all of the above	all of the above
45	The method of mushroom harvesting is	handpicking	machine lifting	lifting with trowels	using scissors	handpicking

Unit III

Equipment and sterilization techniques. Isolation and culture of spores, culture media preparation. Production of mother spawn, multiplication of spawn—Inoculation technique—Cultivation technology—Substrates, composting technology, bed, polythene bag preparation, spawning—casing—cropping—Mushroom production—harvest—packing, storage and marketing.

Tissue Culture

Tissue culture technique is used to bring the edible mushroom to pure cultures so that the mushroom fungus can further be used to prepare spawn, which is an essential material for mushroom cultivation. This nucleus culture is grown on Potato Dextrose Agar medium in test tubes. A small tissue from a well-grown mushroom is aseptically transferred to agar medium in a test tube in a culture room.

The test tubes are incubated under room temperature for 10 days for full white growth of fungal culture. This is further used for preparation of mother spawn.

Procedure

1. Select well-grown, disease-free button mushroom early in the morning and keep it on a clean paper for 2-3 hr, to get certain amount of moisture present in the mushroom to get evaporated.
2. Clean the culture room/laminar flow chamber with antiseptic solution.
3. Keep the sterilized PDA slants, razor blades, forceps etc. inside the chamber and put on the UV light.
4. After 20 minutes, put off the UV light and start working after 5 minutes.
5. Sterilize all the instruments to be used by exposing to Bunsen burner.
6. Take in the mushroom and split open the mushroom longitudinally into two halves.
7. Using a blade cut a small piece of tissue from the centre of the split mushroom at the junction of pileus and stipe.
8. Remove the cotton plug of the agar slant and the tissue is aseptically placed inside the slant by using a sterilized forceps and close it immediately.
9. After transferring tissues from the mushroom, the tubes are arranged in a wire basket and kept in a clean room at room temperature for the growth of the fungus.
10. Observe the tube at periodical intervals and remove the contaminated ones. The tubes will be ready for further use within another ten days. The base spawn is used for preparation of mother spawn.

Precautions to be observed:

- Wash the hands with antiseptic lotion before starting work inside the chamber. If possible, it is better to use hand gloves while operation.
- It is better that the maximum of two persons may work inside the room at a time. Avoid unnecessary talking while working inside the room.
- While separating the tissue from the centre of the mushroom it should not touch the bottom or sides of the mushroom.

Expt: 3 Spawn making using sorghum

Mother Spawn:

Mother spawn is nothing but the mushroom fungus grown on a grain based medium. Among these several substrate materials tested by TNAU, Coimbatore, sorghum grains are the best substrate for excellent growth of the fungus. Well-filled, disease-free sorghum grains are used as substrate for growing the spawn materials. The various steps involving in preparation of mother spawn are listed below here under.

Procedure

1. Wash the sorghum grains in water thoroughly to remove chaff and damaged grains.
2. Cook the grains in an autoclave/vessel for 30 minutes just to soften them.
3. Take out the cooked grains and spread evenly over a Hessian cloth on a platform to remove the excess water.
4. Mix Calcium carbonate (CaCO₃) thoroughly with the cooked, dried grains @ 20g/kg.
5. Fill the grains in polypropylene bags up to 3/4th height (approximately 300-330g/bag), insert a PVC ring, bold the edges of the bag down and plug the mouth tightly with non-absorbent cotton wool.
6. Cover the cotton plug with a piece of waste paper and tie tightly around the neck with a jute thread.
7. Arrange the bags inside an autoclave and sterilize under 20 lbs. pressure for 2 hours.
8. Take out the bags after cooling and keep them inside the culture room and put on the UV light.

9. After 20 minutes put off the UV light and start working in the culture room. Cut the fungal culture into two equal halves using an inoculation needle and transfer one half portion to a bag. Similarly, transfer another half portion of the culture to another bag.
10. Incubate the inoculated bags in a clean room under room temperature for 10 days for further use to prepare bedspawn. What is mushroom spawn? Is it a seed or a root? Do you plant it or sow it, or how do you prepare it? Are some of the questions asked men and again. To the general public there seem to be some great mystery surrounding this spawn question; in fact, it appears to be the chief enigma connected with mushroom-growing. Now, the truth is, there is no mystery at all about the matter. What practical mushroom growers call spawn, botanists term mycelium.

The spawn is the true mushroom plant and permeates the ground, manure, or other material in which it may be growing; and what we know as mushrooms is the fruit of the mushroom plant. The spawn is represented by a delicate white mold-like network of whitish threads which traverse the soil or manure. Under favorable circumstances it grows and spreads rapidly, and in due time produces fruit, or mushrooms as we call them. The mushrooms bear myriads of spores which are analogousto seeds, and these spores become diffused in the atmosphere and fall upon the ground. It is reasonable to suppose that they are the origin of the spawn which produces the natural mushrooms in the fields, also the spawn we find in manure heaps. But we never have been able to produce spawn artificially from spores, or in other words, mushrooms have never been grown by man, so far as I can find any authentic record, from "seed." How, then, do we get the spawn? By propagation by division. We take the mushroom plant or [Pg 79] spawn, as we call it, and break it up into pieces, and plant these pieces separately in a prepared bed of manure or other material, under conditions favorable for their growth, and we find that these pieces of spawn develop into vigorous plants that bear fruit (mushrooms) in about two months from planting time. When the spawn has borne its full crop of fruit it dies.

Well, then, if we cannot produce spawn from spores, and the spawn in the beds that have borne mushrooms has died out, how are we to get the spawn for our future crops? is a question that may suggest itself to the inexperienced. By securing it when it is in its most vigorous condition, which is before it begins to show signs of forming mushrooms, and drying it, and keeping it dry till required for use. But in order to secure the spawn we need to take and keep with it the manure to which it adheres or in which it is spreading. In this way it can be kept in good condition for several years and without its vitality being perceptibly impaired. Keeping it dry merely suspends its growth; as soon as it is again submitted to favorable conditions of moisture and heat its pristine activity returns.

Mushroom spawn can be obtained at any seed store. Our seedsmen always keep it in stock, both the brick (English), and the flake (French) spawn. It is retailed in quantities of one pound or more, and as the article is perfectly dry it can be easily sent by mail in small quantities.

These seedsmen import it from Europe every year along with their seeds. A prominent Boston seedsmen writes me: "We get our supply through the London wholesale seedsmen, for the sake of convenience and cheaper ocean

freight, etc. Coming with shipment of other goods and on same bill of lading brings the freight charges down. The low price at which mushroom spawn is sold in quantity can only be maintained with [Pg80] low freight rates, as there is a duty here of 20% on the article."



Fig.21. Brick Spawn.

By direct inquiry of the leading importers in different cities I find that we import about 4500 lbs of French or flake spawn, and 4000 bushels, or 64,000 lbs of English or brick spawn, and that fully a half of this whole importation is handled by these seedmen of New York city. In New York one firm alone, who make a specialty of supplying market gardeners, has in one year imported 1500 bushels of brick spawn. But the vicinity of New York is the great mushroom-growing center of the country, also the best market for mushrooms in the country. One gardener at Jamaica, L.I., bought 1000 lbs of brick spawn at one time, and a neighbor of his bought 400 lbs; this shows what a large quantity of spawn market gardeners require. And the demand this year is unprecedented; some of our leading importers had sold out their supply before the first of November. And it is not private growers so much as market growers [Pg81] who are the cause of this; the marketmen find there is money in growing mushrooms and they are going into it.

Spawn comes in the form of dry, hard, solid manure bricks, and also in the form of flakes or half-frosted straw manure. These bricks and flakes are completely permeated with the mushroom mycelium.

The brick spawn is commonly known as English spawn, and what is imported into this country is made in England, mostly about London. The bricks made by the different manufacturers vary a little in size and weight; in some cases ten bricks go to the bushel, in others fourteen, and in others sixteen. This last is the commonest sized brick, and weighs exactly a pound, and measures about eight and one-half inches long, five and one-fourth inches wide,

and one and one-fourth inches thick; it is what the London spawn makers call a 9x6x2 inch brick, but it shrinks in drying. In retailing brick spawn in this country it is sold by weight and not by measure.

Mill-track mushroom spawn is advertised by some of our seedsmen, but what they sell under this name is only the ordinary English brick spawn. One of our prominent seed firms who advertise it write me: "Genuine mill-track spawn used to be the best in England, but it has been superseded, although European gardeners still call for English spawn under the name of 'mill-track.'" The real mill-track spawn is the natural spawn that has spread through the thoroughly amalgamated horse droppings in mill-tracks or the cleanings from mill-tracks. It is usually sold in large, irregular, somewhat soft lumps, and is much esteemed by spawn makers for impregnating their bricks, but nowadays, that horses have given place to steam as a motive power in mills, we have no further supply of mill-track spawn for use in spawning our mushroom beds. We do not feel this loss, however, as the spawn now manufactured by our best [Pg82] makers will produce as good a crop of mushrooms as the old mill-track natural spawn used to do.

The flake spawn is what is generally known as French spawn, and is imported into this country from France. But the manufacture of "French" spawn for sale, however, is not strictly confined to France. It is put up in two ways, namely, nicely packed in thin wooden boxes, each containing two or three pounds of spawn, and also loose in bulk when it is sold by weight or measure.

Virgin spawn is what we call natural spawn or wild spawn; that is, the spawn that occurs naturally in the fields, in manure piles, or elsewhere, and without any artificial aid. It is supposed to be produced directly from the mushroom spores, and is not a new growth of surviving parts of old spawn that may have lived over in the ground. It is far more vigorous than "made" spawn, and spawn makers always endeavor to get it to use in spawning the artificial spawn. It is seldom used for spawning mushroom beds because it is not easy to obtain. Now and again we come upon a lot of it in a manure pile; it looks like a netted mass of white strings traversing the manure. As soon as discovered securely you can find, bring it indoors to a loft, shed, or room, and spread it out to dry; after drying it thoroughly keep it [Pg83] dry and preserve and use it as you would French spawn, for it is the best kind of flake spawn. In using virgin spawn for spawning beds I have obtained larger and heavier mushrooms than from "made" spawn, and the beds lasted longer in good bearing, but the weight of the whole crop has not been more than from artificial spawn.

How to Keep Spawn.—Spawn should be kept in a dry, airy place, somewhat dark, if convenient, and in a temperature between 35° and 65°. Wherever things will "must," as in a cellar, cupboard against a wall, or in a close, damp building, is a very poor place for keeping spawn. If the spawn is perfectly dry and kept in a dry, airy place, and not in large bulk, and covered, it will bear a high temperature with apparent impunity, but whenever dampness, even of the atmosphere, is coupled with heat, the mycelium begins to grow, and this, in the storeroom, is ruinous to the spawn. Judging from our natural mushroom crops, the spawn for which must be alive in the ground in winter, one concludes that frost should not be injurious to the artificial spawn, still my experience is that hard frost destroys the vitality of both brick and flake spawn. And this is one reason why I get our full supply of spawn in the fall and keep it myself rather than submit it to the mercy of the seed store.

New Versus Old Spawn.—How long spawn may be kept without its vitality becoming impaired is an unsettled question, but there is no doubt, if properly kept, it will remain good for several years. But I cannot impress too strongly upon the reader the importance of using fresh spawn. Do not use any old spawn at any price; do not accept

it gratis and ruin your prospect of success by using it. It takes three months from the time when the manure is gathered for the beds until the mushrooms are harvested. Can you, therefore, afford [Pg84] to spend this time, and undergo the care and trouble and expense, and court a failure by using gold spawn? We have risks enough with new spawn, let alone old spawn. I do not use any more old spawn, but I have used it often and long enough to be convinced of its general worthlessness, unless preserved with the greatest care.

How to Distinguish Good from Poor Spawn.—This is a very difficult matter, notwithstanding what people may say to the contrary. If we could positively tell good from bad spawn, we would never use bad spawn, and, therefore, with ordinary care, have very few failures in mushroom-growing; for good spawn is the root of success in this business. Spawn differs very much in its appearance; sometimes the bricks show very little appearance of the presence of spawn, and still are perfectly good; and again, we may get bricks that are pretty well interlaced and clouded with bluish white mold or fine threads, and this, too, is good. When the bricks are freely pervaded with pronounced white threads this is no sign that the spawn is bad. Bricks dried as hard as a board may be perfectly good; so, too, may be those that are comparatively soft. Mushroom spawn should have a decided smell of mushrooms, and whatever cobweb-like mold may be apparent should be of a fresh bluish white color, and the fine threads clear white. Prominent yellowish threads or veins are a sign that the mycelium had started to grow and been killed. Distinct white mold patches on the surface of the bricks indicate the presence of some other fungous parasite on the mushroom mycelium; the absence of any mushroom smell in the spawn indicates its worthlessness and that the mycelium is dead. One familiar with mushroom spawn can tell with considerable certainty "very living" spawn and "very dead" spawn, but I am far from convinced that anyone can decide unhesitatingly in the case of middling or weak spawn.

[Pg85] Mr. S. Henshaw, in Henderson's Handbook of Plants, tells us: "The quality of the spawn may be very easily detected by the mushroom-like smell, ... and I should have no hesitation in picking out good spawn in the dark." Sanguine, surely, but I have tried it and found the test wanting. M. Lachaux says that good spawn shows "an abundance of bluish-white filaments well fitted together, and giving off a strongly marked odor of mushrooms. All those portions which show traces of white or yellow mold or have a floury appearance, should be rejected and destroyed." Mr. Wright says: "A brick may be a mass of moldiness, and yet be quite worthless; and if the mold has a spotted appearance, as if fine white sand had been dredged on and through the mass, it is certain there is no mushroom-growing power there.... If thick threads pass through the mass and there are signs of miniature tubercles on them, then the spawn may be regarded as too far gone.... Clusters of white specks on the spawn denote sterility."

Mr. A. D. Cowan, of New York, who has the reputation of being an excellent judge of mushroom spawn, writes me: "To correctly judge the quality of brick spawn by its appearance requires experience in handling it, and a trained eye which enables one quickly to detect good from bad, fair to middling. As two lots seldom come exactly or nearly alike in appearance, it is hardly possible to give precise rules to follow, excepting the never-failing requisite which the spawn must possess to be good, namely, the moldy appearance on the surface, the more the better, without showing threads. Too many of these to a given space are a sure indication of exhausted vitality, arising generally from the bricks being heaped together when in process of manufacture, before they are sufficiently dried. Healthy bricks are usually of a dusty brown color, and of light weight. [Pg86] Black colored spawn is to be avoided, as a rule, and when the black appearance is very prevalent in a cargo of bricks it is a strong indication that the spawn has not run its course; and as it is not expected to do so after it has reached the hands of the retailer it is economy to cast it aside. Some persons break a brick into several pieces to see how it looks inside. To the experienced eye this is not necessary, or even to lay hands upon it, as the outward moldy appearance is the

best of all evidence of its healthy vitality, and this never exists if the bricks have lost their germinating power, excepting, of course, where they have been kept damp, and the spawn has spent its power, which is detected by the white threads appearing in great quantity."

American-made Spawn.—So far as I have been able to find out by diligent inquiry, mushroom spawn is not made for sale in this country. But I am informed that a few growers do save and use their own flakes spawn. Some of our principal growers, Van Siclen, Gardner, and Henshaw, for instance, in time past attempted to make their own spawn, but with only partial success, and now they confine themselves to the imported article. But this state of affairs cannot long continue. The demand here for fresh mushrooms is so great, the industry of mushroom-growing so important, the price of imported spawn so high, and the quantity of foreign spawn imported annually into this country is so large, that, before long, we hope someone will find it to his advantage to make a specialty of growing mushroom spawn in this country to supply the American market. There is no practical operation in connection with the cultivation of mushrooms so little known or understood by the general grower as the growing (or "making," as it is commonly called) and preserving of mushroom spawn. General cultivators in England and France (outside of the Paris caves) do not make their own [Pg 87] spawn; it is a distinct branch of the business, and carried on by specialists who grow mushrooms for sale in winter, and spawn in summer.

The time and attention required to produce a small quantity of first-class spawn are worth more than the cost of the spawn at the seed store. In order to make spawn profitably we must make it in large quantity, and we need not attempt to make it unless we have good materials and conditions for its proper preparation, and will give it every attention possible for its best development.

Because spawn may be made in America is no reason whatever why the American people will buy it. We must produce, at least, as good an article as the best in Europe before we can find countenance in our home market. It is not the shape of the manure brick, its size, fine finish, hardness, softness, or freshness, that counts in this case; it is the fullness and vitality of the mass of mycelium or mushroom plant that is contained within it.

For the mushroom bed is made up it should, within a few days, warm to a temperature of 110° to 120° . Carefully observe this, and never spawn a bed when the heat is rising, or when it is warmer than 100° , but always when it is on the decline and under 90° . In this there is perfect safety. Have a ground thermometer and keep it plunged in the bed; by pulling it out and looking at it one can know exactly the temperature of the bed. Have a few straight, smooth stakes, like short walking canes, and stick the end of these into the bed, twelve to twenty feet apart; by pulling them out and feeling them with the hand one can tell pretty closely what the temperature of the bed is.

All practical mushroom growers know that if the temperature of a twelve-inch thick bed at seven inches from the surface is 100° , that within an inch of the surface of the bed will only be about 95° indoors, and 85° to 90° out of doors. Also, that when the heat of the manure is on the decline it falls quite rapidly, five, or ten degrees, a day, till it reaches about 75° , and between that and 65° it may rest for weeks.

Some years ago I gave considerable attention to this matter of spawning beds at different temperatures. Spawn planted as soon as the bed was made (five days after spawning the heat in interior of bed ran up to 123°) yielded no mushrooms, the mycelium being killed. The same was the case in all beds where the spawn had been planted before the heat in the bed had attained its maximum [Pg 97] (120° or over). Where the heat in the middle of the bed never reached 115° , the spawn put in when the bed was made, and molded over the same day, yielded a small

crop of mushrooms. A bed in which the heat was declining was spawned at 110°; this bore a very good crop, and at 100° and under to 65° good crops in every case were secured, with several days' delay in bearing in the case of the lowest temperatures. But notwithstanding these facts, my advice to all beginners in mushroom growing is, wait until the heat of the bed is on the decline and fall to at least 90°, before inserting the spawn.

Writing to me about spawning his beds, Mr. Withington, of New Jersey, says: "I believe a bed spawned at 60° to 70°, and kept at 55° after the mushrooms appear, will give better results than one spawned at a higher temperature, say 90° **Preparing the Spawn.**—If brick spawn is used cut up the bricks (standard size) into ten or twelve pieces with a sharp hatchet, and avoid, as much as possible, making many crumbs, as is the case generally when a hammer or mallet is used in breaking the bricks. Extra large pieces of spawn are apt to produce large clumps of mushrooms, but this is not always an advantage, as when many mushrooms grow together in a clump they are apt to be somewhat undersized, and in gathering we cannot pluck them all out clean enough so as not to leave a part of the "root" in the ground to poison the [Pg 98] balance of the clump, in cases where several or many of them spring from one common base.

Inserting the Spawn.—When brick spawn is used plant the lumps about an inch deep under the surface of the manure, and about ten inches apart each way. If the spawn looks very good, and the lumps are large do not plant them quite so close as when the spawn shows less mycelium in it, and the lumps are small. Never use a dibber in planting spawn; simply make a hole in the manure with the fingers, insert the lump and cover it over at once, and as soon as the bed has been planted firm it well all over. Although the lumps are buried only an inch deep under the manure, we have to make a hole three or four inches deep to push the lump into to get it buried.

French or flake spawn is inserted in much the same way and at about the same distance, only, instead of cutting it up into lumps, we merely break it into flake pieces about three inches long by an inch thick, and in planting it in the beds, in place of pushing it into the hole, lay in the flake on its flat side and at once cover it.

Many growers plant spawn a good deal deeper than I do, but I have never found any advantage in deep planting. In moderately warm beds, or beds that are likely to retain their heat for a considerable time, I am satisfied that shallow planting is better than deep planting. When we want to mold over our beds soon after spawning them, shallow planting is to be recommended. But if the beds are only 75° to 78°, before being spawned; then I think deep planting is better than shallow planting, because the genial temperature gives the mycelium a better start in life than would the cooler manure nearer the surface.

If there is any likelihood of the surface manure getting wet from the condensed moisture of the atmosphere, I would again cover over the beds with some hay or [Pg 99] straw, and let it remain on until molding time. And if the bed is a little sluggish,—that is, cool,—this covering will help in keeping it warm. Outside beds should be moldered over in three or four days after spawning; inside beds in eight to ten days.

Steeped Spawn.—As brick spawn is so hard and dry I have tried the effect of steeping it in tepid water before planting; some pieces were merely dipped in the water, and others allowed to soak in the pails one-half, one, five, and ten hours. The effect was prejudicial in every instance and ruinous in the case of the long-soaked pieces.

Flake Spawn.—"This is produced by breaking up the brick spawn into pieces about two inches square and mixing them in a heap of manure that is fermenting gently. After lying in this heap about three weeks it will be found one

mass of spawn, and just in the right condition for running vigorously all through the bed in a very short time.... When flakes spawn is used the appearance of the crop is from two to three weeks earlier than when brick spawn is used."— Mr. Henshaw, in first edition of "Henderson's Handbook of Plants." I have tried this method and given it careful attention, but the results were inferior to those obtained where plain, common brick spawn had been used at once.

In all my practice I have found that any disturbance of the spawn when in active growth which would cause a breaking, exposing, or arresting of the threads of the mycelium has always had a weakening influence upon it. I have transplanted pieces of working spawn from one bed to another, as the French growers do, but am satisfied that I get better crops and larger mushrooms from beds spawned with dry spawn than from beds planted with working spawn from any other beds.

LOAM FOR THE BEDS.

In growing mushrooms we need loam for casing the beds after they are spawned, to top dressing the bearing beds when they first show signs of exhaustion, filling up the cavities in the surface of the beds caused by the removal of the mushroom stumps, and for mixing with manure to form the beds. The selection of soil depends a good deal on what kind of soil we have at hand, or can readily obtain.

The best kind of loam for every purpose in connection with mushroom-growing is rich, fresh, mellow soil, such as florists eagerly seek for potting and other greenhouse purposes. In early fall I get together a pile of fresh sod loam, that is, the top spit from a pasture field, but do not add any manure to it. Of course, while this contains a good deal of grassy sod there is much fine soil among it, and this is what I use for mushrooms. Before using it I break up the sods with a spade or fork, throw aside the very toughest parts of them, and use the finer earthy portion, but always in its rough state, and never sifted. The green, soddy parts that are not too rough are allowed to remain in the soil, for they do no harm whatever, either in arresting the mycelium or checking the mushrooms, and there is no danger that the grass would grow up and smother the mushrooms.

Common loam from an open, well-drained fallow field is good, and, if the soil is naturally rich, excellent for any purpose. But do not take it from the wet parts of the fields. Reject all stones, rough clods, tussocks, and the like. Such loam may be used at once.

[Pg 101] Ordinary garden soil is used more frequently than any other sort, and altogether with highly satisfactory results. The greatest objection I have to it is the amount of insects it is apt to contain on account of its often repeated heavy manurings.

Roadside dirt, whether loamy or gritty, may also be used with good results. If free from weeds, sticks, stones and rough drift, it may be used at once, but it is much better to stack it in a pile or rot for a few months before using.

Sandy soil, such as occurs in the water-shed drifts along the roads and where it has been washed into the fields, is much inferior to stiff and more fibrous earth.

I have used the rich dark colored soil from slopes and dry hollows in woods, and, odd though it may appear, as mushrooms do not naturally grow in woods, with success. But it is not as good as loam from the open field.

Peat soil or swamp muck that has been composted for two or three years has failed to give me good returns. The mushrooms will come up through it all right, but they do not take kindly to it.

Heavy, clayey loam is, in one way, excellent, in another, not so good. So long as we can keep it equably moist without making it muddy it is all right, but if we let it get a little too dry it cracks, and in this way breaks the threads of the spawn and ruins the mushrooms that were fed through them.

Loam Containing Old Manure.—Loam in which there is a good deal of old, undecomposed manure, such as the rich soil of four vegetable gardens, is unqualifiedly condemned by some writers because of the quantity of spurious and noxious fungi it is supposed to produce when used in mushroom beds. But I cannot join in this denunciation because my experience does not justify it. This earth is the only kind used by many market gardeners, as they have no other, and certainly without [Pg 102] apparent injurious effect. When I was connected with the London market gardens, some twenty years ago, Steele, Bagley, Broadbent, and the other large mushroom growers in the Fulham Fields case dalled their beds with the common garden soil—perhaps the most manure-filled soil on the face of the earth—and spurious fungi never troubled them. Indeed, I cannot understand why it should produce baneful crops of toad stools when used in mushroom beds, and not of toad stools when used for other horticultural purposes, as on our carnation benches in greenhouses, in our lettuce or cucumber beds, or in the case of potted plants. True, spurious fungi may appear in the earth on our greenhouse benches or frame beds or mushroom beds at any time and in more or less quantity, but I am convinced that the rich earth of the vegetable garden has no more to do with producing toad stools than has any other good soil, and old manure has far less to do with it than fresh manure.

All practical gardeners know how a hot bed, in spring when the heat is on the decline, are to produce a number of toad stools; and, also, that when the bed is "spent," that is, when the heat is altogether gone, the tendency to bear toad stools has gone too. This peculiarity is more apparent in spring than in fall. All mushroom growers know that spurious fungi, when they appear at all, are most numerous three or two weeks before it is time for the mushrooms to come in sight. The same growth appears in the manure piles out in the yard; a few weeks after the strong heat of the manure has gone a lot of toad stools may be observed on and about the heaps, but on the piles of well-rotted cold manure we seldom find toad stools at all.

The fresh, clean stable manure used in mushroom-growing is not apt to be charged with the spores of pernicious toad stools; their presence is always most marked in the case of mixed manures.

[Pg 103] And there is a current idea that mushrooms will not thrive in beds in which old manure abounds, either in the loam or fermenting material; that it kills the mycelium. This, too, I must refute. I have seen heavy crops of spontaneous mushrooms come up in violet and carnation beds in winter, and where the soil consisted of at least one-fourth of rotted manure well mixed with the earth. In cucumber and lettuce beds the same thing has taken place. And in similar beds that have been planted artificially with spawn, good crops of mushrooms have also been raised, and the mycelium, instead of evading the lumps of old manure in the soil often forms a white web right through them.

EARTHING OVER THE BEDS.

This is an important operation in mushroom-growing, and the one for which loam is indispensable. It consists in covering the manure beds, after they have been spawned, with a coating, or casing as it is more commonly called,

of foam. The spawn spreads in the manure and rises up into the casing, where most of the young mushrooms develop, and all find a firm foothold. The loam also contributes to their sustenance. And it protects the manure, hence the spawn, from sudden fluctuations of temperature, and preserves it from undue wetting or drying.

The best soil to use for this purpose is rich, fibrous, mellow loam, such as is described, page 100.

If the manure is fresh and in good condition and the beds are in a snug cellar or closed mushroom house, I would not case them until the second week after spawning, [Pg 104] say about the eighth or tenth day; but were these same beds in an open, airy shed or other building I would case them over some days earlier, say the fourth or fifth day. A fear is often expressed that when beds are cased within three or four days after being spawned the close exclusion of the manure from the air is apt to raise the heat of the manure in the bed, and thereby destroy the spawn; but I have never known of any truth in this theory, and with well-prepared manure I am satisfied no risk of reheating takes place, at least the thermometer does not indicate it. The great danger of early casing is in killing the spawn by burying it too deep in damp material and before it has begun to run through the manure.

I have conducted several experiments in order to satisfy myself regarding when is the proper time to case the beds, and have found no difference in results between beds that were cased over as soon as they were spawned and others that were not cased over until the fourth, seventh, tenth, or fourteenth day after spawning. The good or bad results in the time of casing depend on the condition of the manure in the beds, the depth at which the spawn has been inserted, the openness or closeness of the place in which the beds are situated, and other cultural conditions. But to delay casing as late as the fifteenth or sixteenth day after spawning is injurious to the crop, because in applying the covering of soil we are sure to break many of the mycelium threads that have by this time so freely permeated the surface of the manure. After the fourth week little white knots may be observed here and there on the spawn threads; these are forming mushrooms, and to delay casing the bed until this time would smother these little pinheads, and greatly mar our prospects of a good crop.

Peter Henderson, in his invaluable work, "Gardening for Profit," has given rise to a deep-seated prejudice [Pg 105] against molding over mushroom beds as soon as they are spawned by telling us that in his first attempt at mushroom-growing he had labored for two years without being able to produce a single mushroom, and all because he molded over his beds with a two-inch casing of foam just as soon as he had spawned them. Then he changed his tactics, and did not mold over the beds until the tenth or twelfth day after spawning, and was rewarded with good crops of mushrooms. Now, notwithstanding Mr. Henderson's experience, it is a fact that many excellent growers spawn and mold their beds the same day, and with success. But Mr. H. has done much good in displaying a rock against which many might be wrecked, so much depends upon other cultural conditions. The old practice of inserting the spawn three or more inches deep into the manure bed and then molding it at once with two inches deep of foam was enough to destroy the most potent spawn; nowadays we barely cover the spawn with the manure, and this is how molding over at once is so successful.

All the preparation necessary is to have the loam in medium dry, mellow condition, well broken up with the spade or digging fork, and freed from sticks, stones, big roots, clods, chunks of old manure, and the like.

Sifting the soil for casing the beds is labor lost. Sifted soil has no advantage over unsifted earth, except when it is to be used for top dressing the bearing beds or filling up the holes in their surface.

The condition of the soil should be mellow but inclined to moist. If wet it can only be used clumsily and spread with difficulty; if dry it can be spread easily but not made firm, and on ridge beds cannot be put on evenly. But when moderately moist it can be spread easily and evenly on flat or rounded surfaces, and made firm and smooth.

How deep the mold shall be put on the beds is also [Pg 106] an unsettled question. Some growers recommend three-fourths of an inch, others one, one and one-half, two, or two and one-half inches, and some of our best growers of fifty or seventy-five years ago were emphatic in asserting three inches as the proper depth, but among recent writers I do not find any who go beyond two and one-half inches. My own experience is in favor of a heavy covering, say one and one-half to two inches. In the case of a thin covering the mushrooms come up all right but their texture is not as solid as it is in the case of a heavy covering, nor do the beds continue as long in bearing; besides, "fogging off" is much more prevalent under thin covering than under heavily covered beds; also, when the coating of foam is heavy a great many more of the "pinheads" develop into full sized mushrooms than in the case of thin molded beds.

Opinions differ as to firming the soil. I am in favor of packing the soil quite firm, and have never seen good mushrooms that could not come through a well-firmed casing of foam, and I never knew of an instance where firm casing stopped or checked the spreading of the mycelium or the development of the mushrooms. In the case of flat beds, — for instance, those made on shelves and floors, — a slightly compacted coating (and this is all Mr. J. G. Gardner uses) may be all right, but in the case of along-side-of-walls, ridge, and other rounded beds I much prefer and always use solidly compacted casings.

Mr. Henshaw has for several years used green sods about two inches thick, put all over the bed, grass side down, and beaten firmly. The advantage of using sods instead of soil, he thinks, is that the young clusters of mushrooms never damp or "fog off" as they are apt to do when soil is used.

I have given this green sods method repeated and careful trials, and am satisfied that it has no advantages, in any way, over common fibrous loam; indeed, it is [Pg 107] not as good. No matter how firmly a sod, having its green side down, may be beaten onto a bed of manure, there is barely any union between the two; the sod merely rests upon the dung, but so closely that the mycelium enters it freely. A slight movement or displacement of the sod after the spawn enters it will break the threads of mycelium between the manure and the sod, and this will destroy the immature mushrooms forming in the sod. This gave me a good deal of trouble. Stepping on the sod would disturb it. A clump of strong mushrooms formed under it sometimes displaces it in forcing their way to the surface.

Sods are only fit for use on flat beds where they can lie solid; on rounded or ridge beds they are too liable to be disturbed. And the trouble and expense of procuring sods are too great to warrant their use, even if they had any advantages.

TOP DRESSING WITH LOAM.

In beds that are in full bearing or a little past their best we often find multitudes of very small or what we call "pinhead" mushrooms, that seem to be sitting right on the top of the loam, or clumps that have been raised a little above the surface by growing in bunches, or what we term "rocks"; now a top dressing of finely sifted fresh loam, about one-fourth to one-half inch thick, spread all over the bed, will help these mushrooms materially without doing any of them harm. But while this top dressing assists all mushrooms that are visible above ground, no matter

how small they may be when the dressing is applied, I am not convinced that it [Pg 108] induces greater fertility in the spawn, or, in other words, induces the spawn to spread further and produce more mushrooms than it would were no top dressing applied. I know that this is contrary to the opinions and writings of many, at the same time it is according to my own observation.

Go over the bed very carefully and pick out every soft or "fogged-off" mushroom, no matter how small it may be, and root out every bit of old mushroom stem or tough spongy material formed by it, and in this way get the bed thoroughly cleaned. Then fill up all the holes caused by pulling the mushrooms or rooting out the old stumps, and when the whole surface is level apply the top dressing evenly all over the face of the bed, avoiding, as much as possible, burying the well advanced mushrooms. While it would be very well to pack the dressing smoothly over the bed, it is impracticable; we may press it gently with the back of the hand on the bare spots between the mushrooms, but we should not do this over the mushrooms, no matter how tiny they may be, else many of the "pinheads" will be injured and cause "fogging off."

But we can firm the dressing to the bed by watering it, which may be done over the whole surface of the bed, and without sparing the mushrooms, large or small. Use clear water and apply it gently through a water-pot rose. I always do this, and have never known it to injure the young mushrooms.

In the case of mushroom beds in which black spots have appeared in the crop, I have found that a top dressing of fine, fresh earth applied evenly all over the bed acts, to a certain extent, as a preventive of further attack, but of course has no effect upon any of the already affected mushrooms, large or small.

THE PROPER TEMPERATURE.

The best temperature at which to keep the mushroom house or cellar is 55° to 57° . But much depends upon the method of growing the esculent; the construction of the house or cellar, and other circumstances. Mushrooms can be successfully grown in buildings in which the temperature may be as low as 20° or as high as 65° . By covering the beds well with hay or other protecting material they can be kept warm, even in sharp frosty weather, as the London market gardeners do with their outdoor beds in winter; but when the temperature in the structure in which the mushrooms are grown averages as high as 70° we cannot hope for success; indeed, 65° is too high.

A high temperature in a close house or cellar is injurious; it hurries in the crop and forces up the mushrooms weak and thin-fleshed and with ungainly, long stems; it soon exhausts the bed. The time when its evil effects are least visible is early in the fall and late in spring when the outside temperature is high, and when the beds are in somewhat airy rather than close quarters. In the Dorset cellars there is a steady difference of about 5° in the temperature between the end next the boiler, which is kept at 60° precisely, and that of the farther end, which registers 55° steadily. There is very little difference in the weight of crop produced on the beds at either end of these cellars, but what little there is is in favor of the cooler end. At 60° the crop begins to come in in six to seven weeks after spawning, lasts for three [Pg 110] to four weeks in heavy bearing and a week or more longer in light bearing, and then it gradually dwindles.

In a temperature of 55° it may be seven weeks after spawning before the mushrooms appear. In a temperature of 50° they may take a few days longer in appearing, but, as a rule, they are firm, heavy, short-stemmed, and perhaps a little furry on top and clammy to the touch, and the beds last in good bearing for two months; indeed, often a

whole winter long. But I have failed to find that the whole crop from a bed in a 45° to 50° temperature was any greater than that of a like bed in a 55° to 57° temperature; it is merely a case of getting in six weeks from the warmer house what it takes ten weeks to get from the cooler one.

In a temperature of 50° it is not necessary to cover the beds to increase their warmth, nor is it needful even in one of 45°, if there is a fair warmth in the body of the bed to keep the spawn working; but if the warmth of the interior of the bed falls under 57°, and the atmospheric temperature under 45°, the bed should be kept warm by covering with hay, straw, matting, or other material, or better still by boxing it over and laying this covering on the outside of the boxing. When cold thicken the covering, when warm lessen it.

WATERING MUSHROOM BEDS.

If the beds get dry they should be watered, for mushrooms will not grow well in dry beds or in a dry atmosphere. Watering is an operation requiring much care. In properly-made beds the manure should remain moist enough from first to last, and whatever dryness is evident should be in the loam casing of the beds and the atmosphere. In all artificially heated mushroom houses the beds and atmosphere are apt to get too dry at one time or another; in underground houses or cellars this is less apparent than in above-ground structures; in shaded north-facing houses dryness is less troublesome than in houses more openly placed.

Endeavor by all fair means to lessen the necessity for watering the beds, but when water is needed never hesitate to give it freely. Mulching the beds and maintaining a moist atmosphere are the best preventives. After the beds are spawned and mold is a good plant cover them with a light coating of straw litter or hay to prevent drying, but this mulching should be removed when it is near time for the young mushrooms to appear. A light sprinkling of water over this mulching every few days, but never enough to reach the soil, assists in preserving enough moisture in the bed under the mulch and also in the atmosphere of the house.

Clean, soft water at a temperature of 80° or 90°; a little warmer or a little colder will not hurt, but do not use water higher than 110°, as it might injure the little pin heads, nor lower than the average temperature of the [Pg 112] house, as it would chill the bed, and this should always be avoided.

Use a small or medium-sized watering pot with a long spout and a fine rose sprinkler. Apply the water in a gentle shower over the bed, mushrooms and all, but never use enough to allow it to settle in pools or run off in little streams. Clean water sprinkled over the mushrooms does not appear to hurt them, but they should never be touched with manure water, as it stains them. Just as soon as the surface of the bed shows signs of dryness give it water, the quantity depending upon the condition of the bed. Never let a bed get very dry before watering it. To thoroughly moisten a very dry bed requires a heavy watering; so much, indeed, that the sudden change might injuriously affect the young mushrooms and spawn. Give enough water at a time to moderately moisten the soil, not to soak it, but never sufficient to pass through the soil into the manure. Clean water only should be used until the beds come into bearing, but after that time manure water may be employed with advantage; however, this is not at all imperative; indeed, excellent crops can be and are continually being produced without the aid of manure water at all.

In the case of beds in full bearing, manure water is beneficial to the crop. Apply it from a small watering pot with a long narrow spout but no rose, and pour the liquid on gently over the surface of the bed, running it freely between the clumps but never touching any of the mushrooms. For this reason a rose should not be used.

I have always used manure water for mushrooms more or less, but during the past two seasons—'87-'88 and '88-'89—I have experimented with it continuously and very carefully, using it in some form or other on part of every bed, and am satisfied that manure water made [Pg 113] from fresh horse droppings is the best, and the dark colored liquid, the drainings from manure piles, is the poorest; in fact, this latter is not as good as plain water, for it seems to have a deadening rather than quickening effect upon the beds. Cow manure and sheep manure make a good liquid manure, but still I prefer the horse manure, and although having given hen and pigeon manure and guano of fair tests I am not satisfied that they have benefited the crop, and there is always a risk in their use. Liquid manure made from the contents of the barnyard tank has not done much good, but fresh urine from the horse and cow stables diluted two to five times its bulk has given favorable results.

Mushrooms not only bear with impunity but appear to enjoy a stronger liquid manure more than do any other cultivated plants, and I am satisfied that the weak liquids usually recommended for pot and garden plants would be barely more efficacious than plain water for mushrooms.

The manure water that has given me most satisfaction is prepared as follows: Dump two bushels of fresh horse droppings into a forty-five gallon barrel and fill up with water; stir it up well and let it settle overnight. Drain off the liquid the next day and add a pound of salt peter to it. For use, to a pailful of this liquid add a pailful of warm water. Water of about 80° to 90° is best for mushroom beds. Salt peter is an excellent fertilizer for mushrooms. I use it in two ways, namely: First, powdered and mixed in the soil for casing the beds, at the rate of two ounces of salt peter to the bushel of earth. Second, dissolved in water at the rate of two ounces of salt peter to eight gallons of water, and sprinkled over the beds.

Common salt I use as an insecticide and also as a fertilizer, and am satisfied that it proves beneficial in both ways. Sometimes I sprinkle it broadcast on the surface [Pg 114] of the beds, always on the bare places, never touching the mushrooms, and leave it there for a day or two, then with a fine, gentle sprinkling of water wash it into the soil. This is to help destroy the *anguillulae*. As a fertilizer only dissolve four ounces of salt in ten gallons of water, and with this sprinkle the beds.

A too dry atmosphere can be remedied by sprinkling the floors, walls, or litter coverings on the beds with water, not heavily or copiously, but gently and only enough to wet the surfaces; better to moisten in this way frequently than drench the place at any one time. But I very much dislike sprinkling the beds in order to moisten the atmosphere. An experienced man can tell in a moment whether or not the atmosphere of the mushroom house is too dry. The air in the mushroom houses should always feel moist, at the same time not raw or chilly, and the floor and wall surfaces should present a slow tendency to dry up, and the earth on the beds should retain its dark, moist appearance. The least tendency to dryness should at once be relieved by damping the wall and floor surfaces.

In houses heated by smoke flues, or still more by ordinary stoves and sheet iron pipes, it may be necessary to dampen the floors and walls once or several times a day to maintain a sufficiently moist atmosphere, but where hot water pipes are used and the houses are tight enough to require but little artificial heat, such frequent sprinkling will not be necessary. In the case of beds in unheated structures the ordinary atmosphere is generally moist enough.

Manure Steam for Moistening the Atmosphere.—The late James Barnes, of England, a grand old gardener, writing in the *London Garden*, Vol. III, page 486, describes his method of growing mushrooms sixty years ago, and says: "In winter a nice moist heat was maintained by placing hot stable manure inside, and often [Pg

115] turning it over." Mr. John G. Gardner, of Jobstown, N.J., is one of Mr. Barnes's old pupils and a most successful mushroom grower, and he now practices this same method of moistening the atmosphere by hot manure steam.

In damping the floor of the mushroom house, as well as the beds, I use a medium-sized watering pot and fine rose; but in sprinkling the walls and other parts not readily accessible by the watering pot I use a common garden syringe. **GATHERING AND MARKETING MUSHROOMS.**

This is an important point in the cultivation of this esculent, and should be attended to with painstaking discretion.

When mushrooms are fit to pick depends upon several conditions; for instance, whether for market or for home use, and if for the latter, whether they are wanted for soups or stews. For fresh and attractive appearance and best appreciation in the market, pick them when they are plump and fresh and just before the frill connecting the cap with the stem breaks apart. The French mushrooms should always be gathered before the frill bursts; the English mushrooms also look best when gathered at this time, but they are admissible if gathered when the frill begins to burst and before the cap has opened out flat. If the mushrooms display a tendency to produce long stems pick them somewhat earlier, soon enough to get them with short shanks, for long stems are disliked in market; so, too, are dark or discolored or old mushrooms of any sort. Sometimes we [Pg 116] may not have enough mushrooms ready at one gathering to make it worthwhile sending them to market, and are tempted to let them stay until gathered until to-morrow, when they have grown larger and many more shall have grown big enough to gather. This should never be done. It will give an unfavored, unequal lot, some big, some little, some old, some young. Far better pick every one the moment it is ready to gather, and keep all safe in a cool place and covered until some more are ready for use, and in this way have a uniform appearing lot of young produce.

Mushrooms for soups should always be gathered before they burst their gills; indeed, they are mostly gathered when in a button state; that is, when they are about the size of marbles. In this condition, when cooked, they retain their white appearance and do not discolor the soup. Immature mushrooms are deficient in flavor.

For home use, for baking, stewing, broiling, or for cooking in any way in which the tenderness of the flesh and the delicious aroma of the mushrooms are desirable in their finest condition, let the mushrooms attain their full size and burst their frills, as seen in Fig. 24, and gather them before the caps open out flat, or the gills lose any of their bright pink color. If you let them get old enough for the gills to turn brown before gathering, the mushrooms will become leathery in texture, and lose in flavor and darkens sadly in cooking.



Fig.24. A Perfect Mushroom.

In picking, always pull the mushrooms out by the [Pg 117] root, and never, if practicable to avoid it, cut them over with a knife. In gathering, take hold of the mushrooms and give them a sharp but gentle twist, pressing them down at the same time, and they generally part from the bed without any trouble; then place them in the baskets, root-end down, so as to keep them perfectly clean and free from grit. Sometimes when several mushrooms are joined together in one root-stock and it is impossible to remove one without disturbing the whole, cut it over rather than pull it out. In the case of clumps of young mushrooms, where one cannot be pulled out without displacing some of the others also, cut it out rather than pull it. There is a knack in pulling mushrooms, easily attained by practice. And even when they come up in thick bunches and it would appear impossible to pull out the full-grown ones without disturbing the others, a practiced hand will give them a witch and a pull—they often part from the bed by the gentlest touch—and get them out without unfastening any of the multitude of small buttons that may be growing around them.

The advantages of pulling over cutting are several: It benefits the bed. If we cut over a mushroom and leave its stump in the ground, in a few days decay sets in and a fluffy or spongy substance grows around the old butt, which destroys many of the little mushrooms around it, as well as every thread of mycelium that comes in contact with it. One should be particular to scoop out these stumps with a knife before this condition takes place, and go over the beds every few days to fill up the holes, made in scooping out the old stumps, with fresh loam.

Pulled mushrooms always keep fresh longer than do those that have been cut. In the interest of the market grower they have another advantage. Mushrooms are bought and sold by weight, and as the stems are always [Pg 118] retained to the caps all are weighed together; if part of the stems had been cut off the weight would have been

reduced, and, in like proportion, the price; but if the stems are retained, it is not only are the mushrooms benefited, but the weight, and with it the price, is also increased.

Gathering Field or Wild Mushrooms.—Go in search of them in the morning before the sunshine gets warm and they become too open or old. If you wish to gather and preserve them in their most perfect condition pull them up by the "roots," carefully remove any soil from them, and then lay them orderly in the basket, the root end down; and by spreading a stout sheet of paper over the layer, another may be arranged above it in the same way, and so on until the basket is full. But if you are not so particular and wish them for immediate use, or for ketchup or drying, the common way of cutting them off and carrying them home in bulk will answer well enough.

Marketing Mushrooms.—Most market growers who live immediately around New York City sell direct, and deliver their mushrooms to hotels, restaurants, and fancy fruiterers. But some of them, also most of those who live at a considerable distance from the city, sell their mushrooms through commission merchants in New York; they, in turn, sell in quantities to suit customers.

Mushrooms are sold by the pound, and come into market in boxes made of strong undressed paper. Some growers have light wooden boxes made that hold from one to four pounds of mushrooms each, and these make convenient and strong packages for shipping by express. They may be sent singly, or, as is the case with the paper boxes, several packed together in crates or boxes. In sending directly to hotels, cheap baskets, holding one or several pounds—Mr. Gardner's baskets hold twelve pounds—are often used, but in sending to commission [Pg 119] merchants, who have to deal them out in quantities to suit customers, mushrooms should always be packed in one, two, three or four pound boxes or baskets, preferably one pound. Mushrooms are not like potatoes or apples, that can be handled, re-measured, and repacked without damaging them. Each re-handling will certainly discolor and perhaps break a good many of them, rendering them unsalable, if not worthless.

The utmost care in gathering and packing of mushrooms for shipping is of primary importance. Gather them the moment they are in best condition, no matter whether or not they are to be packed and shipped the same day; never let them blow open before gathering them; and never cut off short stems. Long stems have to be shortened, but not until everything is ready to pack them. With a very soft hair brush dust off any earth that may stick to the cap of the mushroom, and with a harder brush or the back of a knife rub the earth off of the root end of the stem. Then sort the mushrooms,—the big ones by themselves, the middle-sized by themselves, the small or button-sized ones by themselves, and pack each kind by itself. Pack very firmly without bruising, and so as to show the pretty caps to the best advantage. Never pack mushrooms more than two deep without using plenty of soft paper between the layers, and never put a heavy bulk of them into one box or basket. They discolor so easily that, all things considered, about a pound is enough in a box, if we wish them to carry safely and retain their bright, fresh skin without tarnishing.

Mr. Barter, of London, writes me: "The punnets we use for marketing our mushrooms in are the same that are used for strawberries or peaches. These hold just one pound, but it is becoming more customary now to have little boxes made holding from three to five pounds, as these are better for packing in larger cases for long journeys."

MUSHROOM CULTIVATION-17MBU601

Possible questions

1. Differentiate edible mushroom from poisonous mushroom.
 2. Write about *Volvariella volvacea*.
 3. How to prepare bed for mushroom cultivation?
 4. Write the hematological value of mushroom.
 5. Write the processing of mushroom.
 6. Explain the harvesting technique, storage & packaging of mushrooms
 7. Write in detail about the production of mother spawn & its multiplication.
 8. Explain the nutritional values- vitamins & minerals, calorific values of mushroom.
-

S.N	unit-3	Option A	Option B	Option C	Option D	Answer
1	Mushroom fungus can be isolated from _____	Soil	pasture	dead matter	all of the above	all of the above
2	The first step of spawn production was performed on _____	Natural media	Artificial media	other sources	none of the above	Artificial media
3	Which species is used for the grain spawn?	<i>Agaricus spp</i>	<i>Volvariella spp</i>	<i>Pleurotus spp</i>	Both a and b	Both a and b
4	What are the different kinds of grains used	Wheat	Millet	sorghum	all of the above	all of the above
5	Which is used to encourage for the good growth for spawn?	Calcium carbonate	calcium chloride	calcium sulphate	calcium sulphide	calcium carbonate
6	Avoid _____ plugging of spawn bottles	loose	tighten	over tighten	none of the above	loose
7	_____ should be advised once in the month.	fumigation	sterilization	aeration	disinfection	fumigation
8	We should maintain the mushrooms in _____ condition.	good hygiene	poor hygiene	improper handling	none of the above	good hygiene
9	In culture inoculation which is mixed in boiled grains?	chalk powder	gypsum	lime stone	both a and b	Both a and b
10	_____ Cultures can be stored on compost for two years.	Agaricus	bacteria	actinomycetes	molds	agaricus
11	Recently a new method of growing mushroom in	India	Taiwan	China	U.K	Taiwan

	plastic bags has been developed in _____.					
12	For spawning the bag _____ is commonly used.	Molasses	saw dust	wood powder	husk	saw dust
13	In _____ the cultivation of straw mushroom is conducted outdoors	Thailand	Modern cotton waste	Malaysia	Hongkong	Hongkong
14	In _____ method straw is soaked first of all and is mixed with cattle manure before making the stack.	Thailand	Modern cotton waste	Malaysia	Hongkong	Malaysia
15	In _____ method dry rice straw and cotton waste are used to prepare substrate.	Thailand	Modern cotton waste	Malaysia	Hongkong	Modern cotton waste
16	_____ Mushroom grows in nature on the dead wood of a number of hard wood trees.	button	shittake	paddy straw	oyster	shittake
17	_____ Mushroom grows in nature on the dead wood of a number of hard wood trees.	button	shittake	lentinus edodes	oyster	lentinus edodes
18	The mycelium of _____ grows both on composted and fresh straw.	calocybe indica	button	shittake	lentinus edodes	calocybe indica
19	Pasteurization of substrate gives _____ results.	good	bad	moderate	worst	good
20	Wheat grains free from diseases and insect pests should be boiled for _____	10 – 15 mins	15-20min	20-25 min	20-30min	15 – 20 mins

21	The spawned trays are kept in cropping rooms for _____ days for mycelia growth and spread	8 – 10	12 – 15	2 – 6	6 – 9	8 – 10
22	<i>Flammulina velutipes</i> forms _____ fruit bodies.	Small	big	medium	large	small
23	<i>Flammulina velutipes</i> mushroom usually appears in winter season in _____	Japan	India	Malaysia	Hongkong	Japan
24	<i>Flammulina velutipes</i> mushroom is also called as _____ mushroom	winter	summer	rainy	autumn	winter
25	The whole cultivation process from spawning to cropping usually takes about _____ months.	9	6	4	3	3
26	Mushroom is cultivated on logs or on the mixture of saw dust supplement with _____ bran	Wheat	barley	rice	corn	wheat
27	Temperature between ____ to ____ deg C is suitable for cultivation of mushroom	4 – 9	8 – 15	2 – 6	6 – 9	8 – 15
28	Temperature below ____ deg C also affects the formation of fruiting bodies.	2	4	6	8	8
29	_____ is the most versatile method of preserving large quantities of mushroom.	Blanching	freezing	pickling	both a and b	both a and b
30	_____ Mushrooms are taken directly from a cold water basin or sink where they have been cleaned	washed	dried	freezed	none of the above	washed

31	_____ stops enzymatic actions	Blanching	freezing	pickling	both a and b	Blanching
32	The age old method of _____ mushrooms is still one of the best methods of preserving mushrooms.	drying	Blanching	freezing	pickling	drying
33	Drying mushrooms preserves for very _____ period of time.	limited	long	short	none of the above	long
34	the first steps in spawn production are performed on _____.	artificial	natural	superficial	none of the above	artificial
35	_____ Cultures are then made for this starter culture	agar	broth	samples	specimens	agar
36	The _____ culture should be produced from tissue culture.	agar	broth	mother	specimens	mother
37	_____ Spawn is used to inoculate	agar	broth	mother	specimens	mother
38	Good spawn shows _____ mycelial growth	Vigorous	less	high	medium	Vigorous
39	For spawning the bag _____ is commonly used.	Molasses	Saw dust	Wood powder	Husk	Saw dust
40	In _____ method dry rice straw and cotton waste are used to prepare substrate.	Thailand	Modern cotton waste	Malaysia	Hongkong	Thailand
41	Secondary mycelium of mushroom produces umbrella like structure called as	pileus	gills	primary mycelium	tertiary mycelium	pileus
42	Agaricus bisporus was first cultivated in	M.P	H.P	U.P	PUNJAB	H.P

43	In India only three genera of mushroom are considered as edible varieties	Agaricus, as Lentinus, Plammutina	Agaricus, Pleurotus, Volvariella	Pleurotus, Lycoperdon, Geastrum	none of the above	Agaricus, Pleurotus, Volvariella
44	Mushroom are used in the preparation of	pizza	soup	pickles	all of the above	all of the above
45	Mushrooms belong to class	phytomycetes	basidiomycetes	chloromycetes	none of the above	basidiomycetes

Unit IV

Nutritional profile of Mushrooms: protein, amino acids, calorific values, carbohydrates, fats, vitamins & minerals. Medicinal Properties of Mushrooms: Antibacterial, antifungal, antiviral, anti-tumour effect and hematological value. Cardiovascular and renal effect, in therapeutic diets, adolescence, for aged persons and diabetes mellitus. Mushroom nutraceuticals.

Medicinal properties

Main article: Medicinal mushrooms



Ganoderma lingzhi

Some mushrooms are used or studied as possible treatments for diseases, particularly their extracts, including polysaccharides, glycoproteins and proteoglycans.^[39] In some countries, extracts of polysaccharide-K, schizophyllan, polysaccharide peptide, or lentinan are government-registered adjuvant cancer therapies,^{[40][41]} even though clinical evidence of efficacy in humans has not been confirmed.^[42]

Historically in traditional Chinese medicine, mushrooms are believed to have medicinal value,^[43] although there is no evidence for such uses.

Other uses



A tinder fungus, *Fomes fomentarius*

Mushrooms can be used for dyeing wool and other natural fibers. The chromophores of mushroom dyes are organic compounds and produce strong and vivid colors, and all colors of the spectrum can be achieved with mushroom dyes. Before the invention of synthetic dyes, mushrooms were the source of many textile dyes.^[44]

Some fungi, types of polypores loosely called mushrooms, have been used as firestarters (known as tinder fungi).

Mushrooms and other fungi play a role in the development of new biological remediation techniques (e.g., using mycorrhizae to spur plant growth) and filtration technologies (e.g. using fungi to lower bacterial levels in contaminated water).^[45]

Nutrition

Mushrooms (brown, Italian)
or Crimini (raw)

Nutritional value per 100g (3.5 oz)

Energy 94kJ (22 kcal)

Carbohydrates 4.3 g **Fat** 0.1 g

Protein 2.5 g

Vitamins **Quantity %DV[†]**

Thiamine (B1) ^{9%}
0.1 mg

Riboflavin (B2) ^{42%}
0.5 mg

Niacin (B3) ^{25%}
3.8 mg

Pantothenic acid (B5) ^{30%}
1.5 mg

Vitamin B6 ^{8%}
0.11 mg

Folate (B9) ^{6%}
25 µg

Vitamin C ^{0%}
0 mg

Vitamin D ^{1%}
3 IU

Minerals **Quantity %DV[†]**

Calcium ^{2%}
18 mg

Iron ^{3%}
0.4 mg

Magnesium ^{3%}
9 mg

Manganese ^{7%}
0.142 mg

Phosphorus ^{17%}
120 mg

Potassium	10%	448 mg
Sodium	0%	6 mg
Zinc	12%	1.1 mg

Other constituents	Quantity
Selenium	26
ugCopper	0.5 mg
Vitamin D(UVexposed)	1276 IU

Full Link to USDA Nutrient Database entry; (exposed to UV light)

•Units

- µg= micrograms
- mg= milligrams
- IU= International units

†Percentages are roughly approximated using US recommendations for adults.

Source: USDA Nutrient Database

Raw brown mushrooms are 92% water, 4% carbohydrates, 2% protein and less than 1% fat. In a 100 gram (3.5 ounce) amount, raw mushrooms provide 22 source (20% or more of the Daily Value, DV) of B vitamins, such as riboflavin, niacin and DV) and), and a moderate source (10-19% DV) of phosphorus, zinc and or no

Vitamin D

The vitamin D content of a mushroom depends on postharvest handling, in particular the unintended exposure to sunlight. The US Department of Agriculture provided evidence that UV-exposed mushrooms contain substantial amounts of vitamin D.^[17] When exposed to ultraviolet (UV) light, even after harvesting,^[18] ergosterol in mushrooms is converted to vitamin D₂,^[19] a process now used intentionally to supply fresh vitamin D mushrooms for the functional food grocery market.^[20] In a comprehensive safety assessment of producing vitamin D in fresh mushrooms, researchers showed that artificial UV light technologies were equally effective for vitamin D production as in mushrooms exposed to natural sunlight, and that UV light has a long record of safe use for production of vitamin D in food.^[20]

Mushrooms as a source of food

Man has been hunting for the wild mushrooms since antiquity (Cooke, 1977). Thousands of years ago, fructifications of higher fungi have been used as a source of food (Mattila et al., 2001) due to their chemical

composition which is attractive from the nutrition point of view. During the early days of civilization, mushrooms were consumed mainly for their palatability and unique flavors (Rai, 1994, 1997). Present use of mushrooms is totally different from traditional because, a lot of research has been done on the chemical composition of mushrooms which revealed that mushrooms can be used as a diet to combat diseases. The early history regarding the use of mushrooms in different countries has been reviewed by number of workers (Buller, 1915; Rolfe and Rolfe, 1925; Singer, 1961; Atkinson, 1961; Bano et al., 1964; Jandaik and Kapoor, 1975; Bano and Rajarathnam, 1982; Abou et al., 1987; Houghton, 1995). The oriental use of mushrooms is older than the European (Lambert, 1938). Rolfe and Rolfe (1925) mentioned that the mushrooms like *Agaricus campestris*, *Morchella esculenta*, *Helvella crispa*, *Hydnum coralloides*, *Hypoxylon vernicosum* and *Polyporus mylitta* were used much earlier in India. Lintzel (1941, 1943) recommended that 100 to 200 g of mushrooms (dry weight) is required to maintain an optimal nutritional balance in a man weighing 70 kg. Bano et al. (1963) determined the nutritive value of *Pleurotus flabellatus* as 0.974% ash, 1.084% crude fibre, 0.105% fat, 90.95% moisture, 0.14% non-protein nitrogen and 2.75% protein. Bano (1976) suggested that food value of mushrooms lies between meat and vegetables. Crisan and Sands (1978) observed that mushrooms in general contain 90% water and 10% dry matter. Moreso, the protein content varies between 27 and 48%. Carbohydrates are less than 60% and lipids are between 2 to 8%. Orgundana and Fagade (1981) indicated that an average mushroom is about 16.5% dry matter out of which 7.4% is crude fibre, 14.6% is crude protein and 4.48% is fat and oil. Gruen and Wong (1982) indicated that edible mushrooms were highly nutritional and compared favourably with meat, egg and milk food sources. Of several thousand mushroom species known worldwide, only around 2000 are considered edible, of which about 20 are cultivated commercially with only 4 to 5 under industrial production (Chang, 1990). There is also a significant difference in the nutrient contents of pileus versus stalks (Latifah et al., 1996; Zakia et al., 1993).

Carbohydrates

The carbohydrate content of mushrooms represents the bulk of fruiting bodies accounting for 50 to 65% on dry weight basis. Free sugars amount to about 11%. Florezak et al. (2004) reported that *Coprinus atramentarius* (Bull.: Fr.) Fr. contain 24% of carbohydrate on dry weight basis. The mannitol, also called as mushroom sugar constitutes about 80% of the total free sugars, hence it is dominant (Tseng and Mau, 1999; Wannet et al., 2000). Mc-Connell and Esselen (1947) reported that a fresh mushroom contains 0.9% mannitol, 0.28% reducing sugar, 0.59% glycogen and 0.91% hemicellulose. Carbohydrates of *Agaricus bisporus* were reported by Crisan and Sands (1978). Raffinose, sucrose, glucose, fructose and xylose are dominant in it. (Singh and Singh, 2002). Water soluble polysaccharides of mushrooms are antitumor (Yoshioka et al., 1975)

Proteins

Protein is an important constituent of dry matter of mushrooms (Aletor, 1995; Alofe et al., 1995; Fasidi and Kadiri, 1990; Florczak and Lasota, 1995; Zrodowski, 1995; Chang and Buswell, 1996). Lintzel (1941) reported the digestibility of mushroom protein to be as high as 72 to 83%. The proximate analysis of mushroom mycelia has been reported by a number of workers (Humfeld, 1948; Humfeld and Sugihara, 1949; Blocket et al., 1953) and that of *Morchella* species by Litchfield et al. (1963). Protein content of mushrooms depends on the composition of the substratum, size of pileus, harvest time and species of mushrooms (Bano and Rajarathnam, 1982). Protein content of the mushrooms has also been reported to vary from flush to flush (Crisan and Sands, 1978). Haddad and Hayes (1978) indicated that protein in *A. bisporus* mycelium ranged from 32 to 42% on the dry weight basis. Abou et al. (1987) found 46.5% protein on dry weight basis in *A. bisporus*. Samajipati (1978) found 30.16, 28.16, 34.7 and 29.16% protein in dried mycelium of *A. campestris*, *Agaricus arvensis*, *M. esculenta* and *Morchella deliciosa* respectively.

Purkayastha and Chandra (1976) found 14 to 27% crude protein on dry weight basis in *A. bisporus*, *Lentinus*

subnudus, *Calocybe indica* and *Volvariella volvacea*. On dry matter basis, the protein content of mushrooms varies between 19/100 and 39/100 g (Weaver et al., 1977; Breene, 1990). In terms of the amount of crude protein, mushrooms rank below animal meats but well above most other foods including milk (Chang, 1980). On a dry weight basis, mushrooms normally contain 19 to 35% proteins as compared to 7.3% in rice, 12.7% in wheat, 38.1% in soybean and 9.4% in corn (Crisan and Sands, 1978; Li and Chang, 1982; Bano and Rajarathnam, 1988). Verma et al. (1987) reported that mushrooms are very useful for vegetarians because they contain some essential amino acids which are found in animal proteins. The digestibility of *Pleurotus* mushroom proteins is as that of plants (90%) whereas that of meat is 99% (Bano and Rajarathnam, 1988). Rai and Saxena (1989a) observed a decrease in the protein content of mushroom on storage. The protein conversion efficiency of edible mushrooms per unit of land and per unit time is far more superior compared to animal sources of protein (Bano and Rajarathnam, 1988). Mushrooms in general have higher protein content than most other vegetables (Bano and Rajarathnam, 1988) and most of the wild plants (Kallman, 1991). Sharma et al. (1988) reported 14.71 to 17.37% and 15.20 to 18.87% protein in the fruiting bodies of *Lactarius deliciosus* and *Lactarius sanguifusus* respectively. Mushrooms contain all the essential amino acids required by an adult (Hayes and Haddad, 1976). Gupta and Sing (1991) reported 41.4% essential amino acids in *Podaxis pistillaris*. Friedman (1996) reported that the total nitrogen content of dry mushrooms is contributed by protein amino acids and also revealed that crude protein is 79% compared with 100% for an ideal protein.

Fats

In mushrooms, the fat content is very low as compared to carbohydrates and proteins. The fats present in mushroom fruiting bodies are dominated by unsaturated fatty acids. Singer (1961) determined the fat content of some mushrooms as 2.04% in *Suillus granulatus*, 3.66% in *Suillus luteus* and 2.32% in *A. campestris*. Huges (1962) observed that mushrooms are rich in linolenic acid which is an essential fatty acid. Total fat content in *A. bisporus* was reported to be 1.66 to 2.2/100 g on dry weight basis (Maggioni et al., 1968). Ogundana and Fagade (1981) indicated that mushrooms have 4.481% fats on dry weight basis. Kanwar et al. (1990) has reported a fat content of 11.52% in the *Amanitaceae* fruiting bodies on dry weight basis. In 100 g fresh matter of *A. bisporus* (Lange) Sing and *Pleurotus ostreatus* (Jacq: Fr.) Kumm, the content of fatty compounds were found to be 0.3 and 0.4 g respectively (Manzi et al., 2001), but on dry weight basis, it is 2 and 1.8 g respectively (Shah et al., 1997). Aletor (1995), Manzi et al. (2001), Sanmeet et al. (2003) and Manzi et al. (2004) worked on the fibre content of different mushrooms. Mushrooms are considered good sources of fats and minerals (Jiskani, 2001). Yilmaz et al. (2006) and Pedneault et al. (2006) reported that fat fraction in mushrooms is mainly composed of unsaturated fatty acids.

Vitamins

Mushrooms are one of the best sources of vitamins especially Vitamin B (Breene, 1990; Mattila et al., 1994; Zrodowski, 1995; Chang and Buswell, 1996; Mattila et al., 2000). Vitamin content of edible mushrooms has been reported by Esselen and Fellers (1946), Block et al. (1953) and Litchfield (1964). Manning (1985) gave a comprehensive data of vitamin content of mushrooms and some vegetables. According to Mattila et al. (1994), wild mushrooms contain much higher amounts of vitamin D₂ than dark cultivated *A. bisporus*. Mushrooms also contain vitamin C in small amounts.

Mineral constituents

The fruiting bodies of mushrooms are characterized by a high level of well assimilated mineral elements. Major mineral constituents in mushrooms are K, P, Na, Ca, Mg and elements like Cu, Zn, Fe, Mo, Cd form minor constituents (Bano and Rajarathnam, 1982; Bano et al., 1981; Chang, 1982). K, P, Na and Mg constitute about 56 to 70% of the total ash content of the mushrooms (Li and Chang, 1982) while potassium alone forms 45% of the total ash. Abou-Heilah et al. (1987) found that content of potassium and sodium in *A. bisporus* was 300 and 28.2

ppm. respectively. *A. bisporus* analysis showed high amount of K, P, Cu and Fe (Anderson and Fellers, 1942). Kaul (1978) reported that *M. esculenta* contains Ca (0.5776 mg), P (3.313 mg), Fe (1.213 mg) and K (3.831 mg). Varo et al. (1980) reported that *A. bisporus* contains Ca (0.04g), Mg (0.16), P (0.75g), Fe (7.8 g), Cu (9.4 mg), Mn (0.833 mg) and Zn (8.6mg) per kilogram fresh weight. Mushrooms have been found to accumulate heavy metals like cadmium, lead, arsenic, copper, nickel, silver, chromium and mercury (Schmitt and Sticher, 1991; Mejstrik and Lepsova, 1993; Wondratschek and Roder, 1993; Kalac and Svoboda, 2000; Svoboda et al., 2001; Issiloglu et al., 2001; Malinowska, 2004). The mineral proportions vary according to the species, age and the diameter of the fruiting body. It also depends upon the type of the substratum (Demirbas, 2001). The mineral content of wild edible mushrooms has been found higher than cultivated ones (Aletor, 1995; Mattila et al., 2001; Rudawska and Leski, 2005).

Medicinal importance

Medical mycology is as old as traditional uses of mushrooms. They have been used in medicines since the Neolithic and Paleolithic eras (Samorini, 2001). First century Greek physician Dioscorides, included the lurch polypore, (*Fomitopsis officinalis* (Villars: Fr.) Bond and Singer, Polyporaceae; syn. *Laricifomes officinalis* (Villars: Fr.) in his *De Materia Medica* known then as *Agaricum* and latter as the Quinine conk. It was used for the treatment of "consumption", a disease now known as tuberculosis. Although mushrooms as medicine have been used in China since 100 A.D. (Gunde, 1999), but it was only in 1960 that scientists investigated the basic active principles of mushrooms which are health promoting. Mushrooms have been used in health care for treating simple and age old common diseases like skin diseases to present day complex and pandemic disease like AIDS. They are reputed to possess anti-allergic, anti-cholesterol, anti-tumor and anti-cancer properties (Jiskani, 2001). Aqueous extracts from *Pleurotus sajorajua* proved good in renal failure (Tam et al., 1986). The first successful research discovered the antitumor effects of the hot water extracts from several mushrooms (Ikekawa et al., 1969). The main components proved to be polysaccharides especially β -D-glucans. Chihara et al. (1969) isolated from the shiitake fruiting bodies, an antitumor polysaccharide, which was named lentinan. Bahl (1983) reported that mushrooms cure epilepsy, wounds, skin diseases, heart ailments, rheumatoid arthritis, cholera besides intermittent fevers, diaphoretic, diarrhea, dysentery, cold, anesthesia, liver disease, gall bladder diseases and used as vermicides. Most of the mushroom drugs are now available in tablet form in China (Yanget al., 1993). In underdeveloped countries where protein malnutrition has taken epidemic proportions, Food and Agricultural Organization (F. A. O.) has recommended mushroom foods to solve the problem of malnutrition (Sohi, 1988). *Mannentake* (*Ganoderma lucidum*) are known to lower blood pressure and serum cholesterol concentration of hypertensive rats (Kabiret al., 1988). *Lentinus tigrinus* and *G. lucidum* are proved anti-cholesterolemic (Renet al., 1989). *Lentinus edodes* has been used to enhance vigour, sexuality, energy and as an anti-aging agent (Gareth, 1990). *Lentinus sulphureus* obtained from *Lentinus* species inhibits HIV (Gareth, 1990). Jonget al. (1991) reported that mushrooms cause regression of the disease state. Mushroom medicines are without side effects (Sagakami et al., 1991). Puffballs have been used in urinary infections (Buswell and Chang, 1993). Maitake extract has been shown to kill HIV and enhance the activity of T-helper cells (Nanba, 1993; King, 1993). *Ganoderma* nutraceuticals have exhibited promising antiviral effects like, anti-hepatitis B (Kino et al., 1989), anti-HIV (Kim et al., 1993; Liu and Chang, 1995). Dreyfuss and Chapela (1994) reported hundreds of secondary metabolites of fungal origin possessing biological activity. Mushrooms act as biological response modifiers by promoting the positive factors and eliminating the negative factors from the human body and thus regarded as the fourth principal form of the conventional cancer treatment (Yanget al., 1993). *G. lucidum* (Fr.) Karst is believed to act as an anti-inflammatory agent (Stavinoha et al., 1991); acts as anti-diabetic (Teow, 1997). It is also used by Indian tribals for treating joint pain (Harshet al., 1993). Hobbs (1995) reported various medicinal uses of mushrooms like reishi, cordyceps, enoki, maitake, lion's mane and splitgill for

cancer treatment; shiitake, blazei, reishi, enoki, cordyceps, maitake, mesima and oyster were found effective against cholesterol reduction. Reishi, cordyceps, shiitake and maitake is used for reducing stress. Lion's mane has been used for memory improvement; reishi for inducing sleep, cordyceps for physical endurance and sexual performance, reishi, cordyceps, chaga and lion's mane for asthma and allergy treatment. Shiitake, cordyceps, chaga, shiitake and turkey tail as liver protectants; reishi, maitake, turkey tail and shiitake for treating diabetes. It is also believed to be a good health elevator (Mizuno, 1996). Auricularia species were used since times for treating hemorrhoids and various stomach ailments (Chang and Buswell, 1996). Pleurotus tuber-regium mushroom have been used for curing headache, high blood pressure, smallpox, asthma, colds and stomach ailments (Oso, 1997; Fasidi and Olorunmaiye, 1994). It has been reported that *P. ostreatus* lowers the serum cholesterol concentration in rats (Bobek et al., 1996). PSK, an anticancer drug from the mushroom, *Coriolus versicolor* accounted for 25.5% of the country's total sales in Japan in 1987 as anticancer drug (Chang and Buswell, 1996). Puffballs (*Clavatia*, *Lycoperdon*) have been used for healing wounds (Delena, 1999). Pharmaceutical substances with potent and unique health enhancing properties have been isolated from mushrooms (Wasser and Weis, 1999). Fresh mushrooms are known to contain both soluble and insoluble fibres; the soluble fibre is mainly beta-glucans polysaccharides and chitosans which are components of the cell walls (Sadler, 2003). Soluble fibre present in mushrooms prevents and manages cardiovascular diseases (Chandalia et al., 2000). Wasser (2005) reported that mushroom health supplements are marketed in the form of powders, capsules or tablets made of dried fruiting bodies, extracts of mycelium with substrate, biomass or extract from liquid fermentation. *P. sajor-caju* has been found to be inductive for growth of probiotic bacteria (Oyetayo et al., 2005). Cordyceps *sinensis* also treated a half caterpillar and half mushroom has been known and used for many centuries in traditional Chinese medicine. Cordyceps has been used to induce restful sleep, acts as anticancer, antiaging, and antiasthma agents besides proved effective for memory improvement and as sexual rejuvenator (Sharma, 2008).

Antioxidant activity

Antioxidants are chemical compounds that protect cells from the damage caused by unstable molecules known as free radicals. Free radicals are powerful oxidants and those chemical entities that contain unpaired electrons. They are capable of randomly damaging all components of the body, viz. lipids, proteins, DNA, sugars and are involved in mutations and cancers (Przybytniak et al., 1999). The nascent oxygen is trapped by enzymes like superoxide dismutase, catalase and glutathione peroxidase. Over production of free radicals creates oxidative stress. The antioxidants are an important defense of the body against free radicals and mushrooms which are rich sources of these antioxidants (Mau et al., 2004; Puttaraju et al., 2006; Ferreira et al., 2007; Oyetayo et al., 2007). Waxycap mushroom extracts (*Hygrocybe coccinea*) are inhibitory to sarcoma (Ohtsuka et al., 1997). Immunocetals isolated from more than 30 mushroom species have shown anticancer action in animals (Wasser and Weis, 1999). Schizophyllan from *Schizophyllum commune* is effective against head and neck cancer (Kimura et al. 1994; Borchers et al., 1999). Antioxidant property of compounds is correlated with their phenolic compounds (Velioglu et al., 1998). Kim and Kim (1999) reported that mushroom extracts possess DNA protecting properties. *G. lucidum* extracts can trap number of free radicals 2602 J. Med. Plant. Res. (Jones and Janardhanan, 2000). Mau et al. (2001) found antioxidant properties of several ear mushrooms. Many species of mushrooms have been found to be highly potent immune enhancers, potentiating animal and human immunity against cancer (Wasser and Weis, 1999; Borchers et al., 1999; Kidd, 2000; Fenget al., 2001). Tyrosinase from *A. bisporus* is antioxidant (Shi et al., 2002). Lakshmi et al. (2005) determined antioxidant activity of *P. sajor-caju*. Russell and Paterson (2006) observed that triterpenoides are the main chemical compounds in *G. lucidum*. Camptothecin is responsible for antioxidant properties in *G. lucidum* (Zhou et al.,

MUSHROOM CULTIVATIONS-17MBU601A

POSSIBLE QUESTIONS

1. Differentiate edible mushroom from poisonous mushroom.
 2. Write about *Volvariella volvacea*.
 3. How to isolate & culture the spores for mushroom cultivation?
 4. Write about the anti-tumour effect of mushroom.
 5. Write the steps involved in post harvest technologies of mushroom cultivation.
 6. Write about the Anti tumour effect & Hematological value of mushroom.
 7. Explain the outline of nutritional profile of mushroom.
 8. Explain the spore germination & life cycle of *Agaricus bisporus*.
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SN	unit-4	Option A	Option B	Option C	Option D	Answer
1	Mushrooms considered as _____ source of food.	alternative	moderate	generalised	all of the above	alternative
2	Mushrooms are rich in _____	fats	carbohydrates	proteins	minerals	proteins
3	_____ is a building material for all body tissues	fats	carbohydrates	proteins	minerals	proteins
4	Proteins are made up of _____	fats	carbohydrates	aminoacids	minerals	aminoacids
5	Mushroom proteins contain _____ essential amino acids.	5	2	9	12	9
6	The moisture content of the fresh mushroom is _____	60%	90%	40%	30%	90%
7	The moisture content of the dried mushroom is _____	5%	15%	3%	6%	15%
8	Mushrooms contains very low level of _____	Carbohydrates	aminoacids	minerals	fats	fats
9	The _____ content of mushrooms are quite low.	Carbohydrates	aminoacids	minerals	fats	Carbohydrates
10	The carbohydrate content in mushroom are _____	high	quitelow	very low	moder	quitelow

					ate	
11	Mushrooms contain _____ level of fats.	high	quitelow	very low	moderate	very low
12	Fresh mushrooms contain _____ level of moisture	high	quitelow	very low	moderate	high
13	There are _____ in calories, carbohydrates and calcium.	high	low	moderate	none of the above	low
14	Which vitamins are present in mushrooms	thiamine	riboflavin	niacin	all of the above	all of the above
15	Mushrooms are highly _____.	Toxic	Nutritious	non toxic	fatty	Nutritious
16	They are _____ in calories, are great sources of fiber and protein (good for plant-based diets	High	low	moderate	none of the above	low
17	They are packed with many _____ as more colourful fruits and vegetables	Antioxidants	Toxic	non toxic	fatty	Antioxidants
18	Eating mushrooms may also help to prevent _____	Respiratory infections	Toxic	non toxic	fatty	Respiratory infections
19	_____ may help you live a longer, healthier life.	Antioxidants	Anti pulmonary	Both a and b	None of the	Both a and b

		nts	infection		above	
20	Mushrooms are a _____.	gut friendly food	highly toxic	poisonous	respiratory infection	gut friendly food
21	They also contain ____ types of dietary fibers, beta-glucans and chitin.	6	8	2	9	2
22	Mushrooms contain a class of proteins called _____ which are able to bind to abnormal cells and cancer cells and label the cells for destruction by our immune system.	Lectins	pectins	albumin	none of the above	lectins
23	Some wild mushrooms are _____ to humans.	Not edible	Toxic	Not edible and are toxic	none of the above	Not edible and are toxic
24	The products of mushrooms are	mushroom cofee	mushroom elixir	mushroom powder	all of the above	all of the above
25	The _____ content in mushrooms all contribute to cardiovascular health	fibre	potassium	vitaminC	all of the above	all of the above
26	Potassium and sodium work together in the body to help regulate	blood pressure	high pressure	low pressure	none of the above	blood pressure
27	One cup of chopped or sliced raw white mushrooms contains	15 calories	10grams of	2.2 grams of	all of the	all of the

			fat	protein	above	above
28	Mushrooms are rich in ____ vitamins	A	B	C	D	B
29	The B vitamins help the body to form _____	RBC	WBC	Both a and b	none of the above	RBC
30	Reishi mushroom contains chemicals that seem to have a variety of _____ effects, including activity against tumors (cancer) and beneficial effects on the immune system	Potentially beneficial	Toxic	non toxic	none of the above	Potentially beneficial
31	Vitamin D (both Vitamin_____) which plays a critical role in bone health, immune system, energy and mood.	D1 and D2	D3 and D4	D2 and D3	D4 and D5	D2 and D3
32	Mushrooms have _____ fiber which helps support digestion and may help increase beneficial gut bacteria in your digestive system.	Dietary	Diet	Both a and b	none of the above	Dietary
33	Mushrooms provide a variety of essential ecosystem functions, including:	Absorbing pollution	Cleaning polluted soil	Producing natural insecticides	all of the above	all of the above
34	Mushrooms provide a variety of essential ecosystem functions, including:	Breaking down nerve agents	Providing a sustainable fuel source	Producing rich soil for farms and forests	all of the above	all of the above
35	Producing rich soil for farms and forests	high	low	moderate	very	very low

					low	
36	Some wild mushrooms are _____ to humans	Not edible	Toxic	Not edible and are toxic	none of the above	Not edible and are toxic
37	The moisture content of the fresh mushroom is _____	60%	90%	30%	40%	90%
38	Mushrooms contains very low level of _____	Carbohydrates	fats	proteins	none of the above	fats
39	In which of the following stages of development mushrooms should be picked	15 to 20 ⁰ C	30 to 35 ⁰ C	not certain	22 to 25⁰C	22 to 25⁰C
40	FYM is	farmyard manure	fungal yeast medium	fy medium	none of the above	farm yard manure
41	Media for mushroom	PDA	NA	KBA	SSA	PDA
42	the characterisation of poison mushroom is	pink spore	bright coloured	hot burning taste	all of the above	all of the above
43	Which mushroom is less poisonous out of the following	Amantia phalloides	Amantia muscara	Russia emitica	all of the above	Russia emitica
44	Mushroom contains ----- amount of protein	100%	10%	negligible	20-40%	20-40%

45	Wild mushrooms grow during	rainy season	october season	summer	anytime	rainy season
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KAHE

Unit V

Problems in cultivation—diseases, pests and nematodes, weed moulds and their management strategies. Mushroom economics: economics of spawn and mushroom, cultivation, postharvest technologies. Processing and preservation of mushrooms. Mushroom research centres in India.

INSECT AND OTHER ENEMIES.

The mushroom grower has his full share of insects to contend with, and in order to overcome them one should acquaint himself with them, and know what they are, what they do, when they came, and how to destroy them. One should study the diseases and mishaps of his crop and endeavor to know their cause. If we know the cause of failing health in plants, even in mushrooms, we can probably stop or devise a remedy for the disease or means to prevent its recurrence, and if we can not benefit the present subject we are forewarned against future attacks. But there is a deal of mysterious trouble in this direction in mushroom-growing. We are likely to know something about the depredations committed by insects or parasitic molds above ground, but I am sure there is a good deal of mischief going on underground of which we know very little, if anything. The ills to which the mycelium is subject are not at all fully understood.

"Maggots."—This is the common name among practical mushroom growers for the larvæ of a species of fly (Diptera) which from April on through the warm summer months renders mushroom-growing unprofitable. It is unavoidable, and so far has proved invincible. It attacks the mushrooms in deep cellars, above-ground houses, greenhouses, or frames, and is often quite common in early appearing crops in the open fields. We sometimes read that it does not occur in unheated cellars, but this is a mistake, for in our unheated tunnel [Pg 123] cellars, where the temperature in April does not exceed 55°, maggots always appear about the end of this month. But it is true that in the case of cool houses and where the beds are covered over with hay or straw maggots do not appear as early in the season as they do in warm houses and open beds. While rigid cleanliness, and care in keeping the house or cellar closed, no doubt have much to do in lessening the trouble, I have never been able to overcome it, and know of none who has. We simply stop growing mushrooms in summer.

The maggots or larvæ are about three-sixteenths to four-sixteenths of an inch long, white with black head, and appear in all parts of the mushroom, but mostly in the cap and at the base of the stem, and perforate hither and thither leaving behind them a disgusting network of burrows. The tiny buttons, about as soon as they appear at the surface of the ground, are infested, but this does not check their growth, and when they become mushrooms large enough for gathering, unless it be for a dark looking puncture or tracing now and then visible on the outside of the caps and stems, there are but few signs to indicate to the inexperienced eye the presence of maggots. And this is why maggots mushrooms are so often found exposed for sale in summer. But in large or full-grown mushrooms, and especially the white-skinned varieties, their presence is visible enough. Although very repugnant, however, and utterly unfit for food, maggot mushrooms are not poisonous.

But all the mushrooms of summer crops are not maggoty, only a large proportion of them; the evil begins in April, and increases as the summer advances, until August, when it decreases, and in October completely stops—at least this is my experience.

A solution of salt, saltpeter, or ammonia sprinkled over the surface of the beds does not, in this case, do any good as an insecticide, pyrethrum powder diffused [Pg 124] through the atmosphere, and tobacco smoke, have been ineffectual. Burning a lamp set in a basin of water with a little kerosene floating on the surface is almost doubtful

operation. Multitudes of flies are destroyed by this lamp trap, but they are the poor little innocent "manure flies," and the atmosphere of the house is vitiated and rendered unhealthy for the crop. I have tried these lamp traps season after season, and never knew of their doing any good; that is, the maggots seemed just as numerous in the lamp-trapped cellar as in the other cellar in which no lamp trap had been used.

Regarding this "maggots" question, Mr. J. F. Barter, of London, writes me: "During the summer months the outdoor mushrooms get maggoty before they are big enough to gather, but of course they can be grown in cool cellars all the year round.... I know of no sure cure for them (the maggots); of course a light sprinkling of salt with manure or mold does prevent, to a certain extent, but it must be used very carefully." Now my experience is, as I have already said, that it is impossible to grow mushrooms here in summer, even in cool cellars, without having them more or less maggoty. As regards the salt and loam preventive, I have tried it lightly and heavily, but without any apparent good effect.

Black Spot.—All mushroom growers are familiar with this disease, but unless it appears in pronounced form very little notice is taken of it, even by market men, for we see spotted mushrooms continually exposed for sale. It appears as dark brown spots, streaks, or freckles, on the top of the mushroom caps, and increases in distinctness and breadth with age. Fig. 25. It is caused by *eel worms (Anguillulæ)*. These minute creatures enter the mushrooms when the latter are in their tiniest pin form and before they emerge from the ground. If a button arises clean it remains clean, if [Pg 125] diseased it continues to be diseased, and it is a fact that if one mushroom in a clump has black spot we usually find that every mushroom in the clump has it. But mushrooms growing from the same bit of spawn and that come up an inch or two away from the spotted ones may be perfectly clean. Black spot has never occurred with my new beds, and seldom in those in vigorous bearing, but it generally appears in beds that have been in bearing condition for some weeks or are declining. It does not confine itself to any particular spot or part of the bed, and sometimes it is much more plentiful than at others. Between October and March we have very little black spot, but as the spring opens this disease increases. During the winter season, with careful attention, perhaps not so much as one per cent will show black spot, but as the warm weather sets in the percentage increases until in May, when as many as twenty per cent may be affected by it.

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Fig. 25. Mushroom affected with Black Spot.

Black spot is a disease, however, that can be controlled. Keep everything in and about the mushroom houses rigidly clean, and as soon as a bed has ceased to bear a crop worth picking clear it out, lime-wash the place it occupied, and make up another bed. Carefully observe that no old loam or manure is allowed to accumulate anywhere, or green scum forms upon the boards, paths, or walls; boiling water impregnated with alum poured over the boards, walls, and other scum-covered surfaces, will kill the eel worms, but it should not be allowed to touch the mushroom beds that are in bearing or coming into bearing. Much can be done to protect the bearing beds from the ravages of this pest: In gathering the mushrooms remove every vestige of old stump and fogged-off mushrooms, keep the holes filled [Pg126] up with fresh loam, and when the bed has been in bearing condition for a fortnight sprinkle it over with a solution of salt, and next day top dress with a half-inch coating of finely sifted fresh loam; firm it to the bed with the back of the hand, for it can not be pressed on with a spade on account of the growing mushrooms.

Is black spot unwholesome? I do not think so. I have never known any ill effects from eating it. The spotted parts are merely flavorless and tasteless. But it is a very disgusting disease, and no one, I am sure, would care to eat eel worms with their mushrooms. Until quite recently I used to regard the black spot as the mark of some parasitic fungus, and, acting under this impression, sent affected mushrooms to Dr. W. G. Farlow, Prof. of Cryptogamic Botany at Harvard University, for his opinion. He wrote me: "I find that the trouble is due to *Anguillulæ*, and I find an abundance of these animals in the brown spots." He advised me to submit them to an expert in "worms." I then sent samples to my kind friend, Mr. William Saunders, of Washington, D.C., who submitted them, for me, to Dr. Thomas Taylor, the microscopist to the U. S. Department of Agriculture, and who replied: "I recommend that you use a sprinkling of scalding water thoroughly over the entire surface of the bed, especially the portion next to the boxing. The scalding water should be applied before the buttons appear, but not penetrate more than one-eighth of an inch below the surface. *Anguillulæ* abound wherever decaying vegetable matter exists.... The green algæ on the outside of flower pots abound in the *anguillulæ*."

Manure Flies.—This is the name we give to the little flies (a species of *Sciara*) that appear in large numbers in spring and summer in our mushroom houses, or, indeed, in hotbeds or structures of any sort where manure is used, as well as about the manure heaps in the [Pg127] yard. On account of their habits they are regarded with much ill-favor. They hop about the house and are continually running over the mushrooms, beds, and walls, in the most suspicious manner. But, notwithstanding this, I am inclined to regard them as perfectly harmless so far as injuring the mushroom crop is concerned, except the fact that they soil the mushrooms somewhat by their traveling over them with their muddy feet.

In attempting to get rid of the maggot fly I have destroyed large numbers of these little innocents, but without any apparent diminution in their numbers. Lachaux recommends: "These flies may be destroyed by placing about a number of pans filled with water to which a few drops of oil of turpentine have been added. The flies are attracted by the odor and drown themselves. They may also be caught with a floating light, in which they will burn their wings and fall into the water." I have found that pure buhach powder dusted into the air or burned on a hot shovel in the mushroom house has been more effective in destroying these flies than either the lamp or drowning process.

Slugs.—These are serious pests in the mushroom house, especially in above-ground structures, and they also occur in annoying numbers in cellars. Wherever hay or straw is used in covering the beds, or there is much woodwork about the house, slugs appear to be most numerous. They are very fond of mushrooms and attack them in all stages, from the tiny button just emerging from the ground to the fully developed plant. In the case of the buttons

or small mushrooms they usually eat out a piece on the top or side of the cap, and as the mushroom advances in growth these wounds spread open and display an ugly scar or disfigurement. They also bite into the stems. But in the case of fresh, full grown mushrooms they seem to have a particular [Pg128] liking for the gills, and eat patches out of them here and there.

"Bullet" or "Shot" Holes.—My attention was first called to these by Mr. A. H. Withington, of New Jersey. They are little holes cut clear through the mushroom caps, as if perforated by a buckshot, and are evidently the work of some insect. He had, before then, submitted some of these perforated mushrooms to Prof. S. Lockwood, who sent them to Prof. C. V. Riley for his opinion. Prof. Riley replied that: "It is quite likely that the damage was done by some myriapod, possibly a Julus, or some of its allies. Only observation on the spot will determine this point." As I never had any trouble with myriapods attacking mushrooms and had seen nothing of this "bullet hole" work in our own beds I was much interested in the question and determined to look out for it, so I marked off a part of a bed and left that uncared for. I soon found out the trouble. These holes are the work of slugs which I have found and watched in the act of eating out the holes. To find the slugs at work, one has to take his lantern and go out and look for them at night. And to find out about plant parasites—be they fungus, or insect—one has to let them alone and watch them. Had we kept up our unsparing hunt for slugs, probably we should not yet have known what caused these "bullet holes," for no slug would have been left alive long enough to eat a hole through a mushroom cap.

Slugs must be caught and killed. We can find them at night by hunting for them by lamp-light; their slimy track glistens and reveals their presence. A few small bits of slate or half-rotten boards with a pinch of bran on them laid here and there about the beds are handy traps; the slugs gather to eat the bran, hide beneath the rotten wood, and can then be caught and killed. Fresh lettuce leaves make a capital trap, but lettuce in [Pg129] January or February are about as scarce as mushrooms themselves. A dressing of salt is distasteful to slugs, and not injurious to mushrooms. Strong, fresh lime water may be freely sprinkled over woodwork, pathways, walls, or elsewhere where slugs might gather and hide themselves; but this solution should not be used upon the mushroom beds. Rigid cleanliness, however, about the mushroom house, and an ever-alert eye for slugs, should keep them under.

Wood Lice.—These are sure to be more or less abundant in every mushroom house, even in the cellars. They crawl in through doors, ventilators, or other interstices, and are brought in with the manure, and find shelter about the woodwork, manure, or any bits of dry litter that may be around. They attack the pinhead and small button mushrooms by biting out little patches in their tops and sides; and although these patches are small to begin with, the blemish spreads as the mushroom grows, and is an objectionable feature. Trapping and killing the insects is the chief remedy. Put part of a half-boiled potato (for which no salt had been used) into a little pasteboard box, and cover the potato with some very dry swamp moss, lay the box on its side, and open at the end on the bed. The wood lice will gather to eat the potato, and remain after feasting because the dry moss affords them a cozy hiding place. Several of these little boxes can be used. Go through the house in the morning, lift the little traps quickly, and shake out any wood lice that may be in them into a tin pail (an old lard pail will do), which should contain a little water and kerosene. These traps may be used for any length of time, merely observing to change the potato now and again to have it in appetizing condition. Hot water or strong kerosene emulsion may be poured about the woodwork, walls, and pathways, to destroy the wood lice, but should not be allowed to touch the beds. Poisoned [Pg130] sweet apples, potatoes, and parsnips have been recommended as baits for these pests, but I must discourage using poison of any sort in the mushroom house. Six or eight inch square pieces of half-rotten very dry boards laid in pairs, one above the other, also make capital traps; the wood lice gather there to hide themselves; these traps should be examined frequently and the insects shaken into the pail containing water and kerosene.

Mites.—Two kinds of mites are very common about mushrooms in spring and summer; one is whitish and smaller than a "red spider" (one of the commonest insect pests among garden plants), and the other is yellowish and as large as or larger than a "red spider." But I do not think that either of these mites is worth considering as a mushroom pest. The yellow mite (probably *Lyroglyphus infestans*) is extremely common in straw litter on the surface of hotbeds, and I have no doubt finds its way into the mushroom house as manure vermin rather than a mushroom parasite. They are the effect and not the cause of injury to the crop. When mushrooms are wounded or cracked, particularly about the stem, the crevices often become abundantly inhabited with these mites, but they do no material damage.

Mice and Rats.—These rodents are very fond of mushrooms, and where they have access to the beds are troublesome and destructive. Both the common house mouse and the white-bellied fence mouse are mushroom destroyers, but, so far, the nimble but timid field mouse (among garden, open air, and frame crops generally) has never yet troubled our mushrooms, but I can not believe that this immunity is voluntary on its part. The mice bite a little piece here and there out of the caps of the young mushrooms, and these bite-marks, as the mushrooms advance in growth, spread open and become unsightly disfigurements. In the case of open mushrooms, however, the mice, like slugs, prefer [Pg131] the gills to the fleshy caps. Rats are far more destructive than mice. Trapping is the only remedy I use, and would not use poison in the mushroom houses for these creatures for obvious reasons. But we should make our houses secure against their inroads.

Toads.—These are recommended as good insect traps to be used in mushroom houses, but I do not want them there; the cure is as bad as the disease. The mushroom bed is a little paradise for the toad. He gets upon it and burrows or elbows out a snug little hole for himself wherever he wishes, and many of them, too, and cares nothing about whether, in his efforts to make himself comfortable, he has heaved out the finest clumps of young mushrooms in the beds.

Fogging Off.—This is one of the commonest ailments peculiar to cultivated mushrooms. It consists in the softening, shriveling, and perishing of part of the young mushrooms, which also usually assume a brownish color. These withered mushrooms do not occur singly here and there over the face of the bed, but in patches; generally all or nearly all of the very small mushrooms in a clump will turn brown and soft, and there is no help for them; they never will recover their plumpness. Some writers attribute fogging off to unfavorable atmospheric conditions,—the temperature may be too cold, or too hot, or the atmosphere too moist, or too dry. I am convinced that fogging off is due to the destruction of the mycelium threads that supported these mushrooms; it is a disease of the "root," to use this expression; the "roots" having been killed, the tops must necessarily perish. If it were caused by unfavorable conditions above ground we should expect all of the crop to be more or less injuriously affected; but this does not occur; the mushrooms in one clump may be withered, and contiguous clumps perfectly healthy.

Anything that will kill the spawn or mycelium threads [Pg132] will cause fogging off to overtake every little mushroom that had been attached to these mycelium threads. Keeping the bed or part of it continuously wet or dry will cause fogging off, so will drip; watering with very cold water is also said to cause it, but this I have not found to be the case. Unfastening the ground by abruptly pulling up the large mushrooms will destroy many of the small mushrooms and pinheads attached to the same clump; and when large mushrooms push up through the soil and displace some of the earth, all the small mushrooms so displaced will probably waste away, as the threads of mycelium to which they were attached for support have been severed. A common reason of fogging off is caused by cutting off the mushrooms in gathering them and leaving the stumps in the ground; in a few days' time these

stumps develop a white fluff or fleck substance, which seems to poison every thread of mycelium leading to it, and all the mushrooms, present and to come, that are attached to this arrested web of mycelium are affected by the poison of the decaying old mushroom stump, and fogg off. Any impure matter in the bed with which the mycelium comes in contact will destroy the spawn and fogg off the young mushrooms. Lachaux complains about the larvæ of two beetles, namely *Aphodius fimetarius* and *Dermestes sellatus*, which "cause great damage by eating the spawn, thereby breaking up the reproductive filaments." Damage of this sort by these or any other insect vermin will cause fogg off. But I have not noticed either of the above beetles or their larvæ about our beds.

Flock.—This is the worst of all mushroom diseases and common wherever mushrooms are grown artificially. It is not a new disease; I have known it for twenty-five years, and it was as common then as it is now, and practical gardeners have always called it *Flock*. I say "worst of all diseases" because I know that mushrooms [Pg 133] affected by it are both unwholesome and indigestible, and I can readily believe that in aggravated cases they are poisonous. It is caused by other fungi which infest the gills and frills of the mushrooms, and render them a hard, flocky mass; sometimes the affected mushrooms preserve their white skin, color, and normal form, at other times the cap becomes more or less distorted. The illustration, Fig. 26, is from life, and a good average of a flock-infested mushroom. In gathering mushrooms the growers should insist that every flock-infested mushroom be discarded, and consumers of mushrooms should familiarize themselves with this disease so as to know and reject every mushroom showing a trace of it.

Flock does not affect all the mushrooms in a bed at any time, and I do not believe it spreads in the bed, or, to use the expression, becomes contagious. If one spot of mildew appears upon a cucumber, rose, or grapevine indoors, and is not checked, it soon becomes general all over the plant or plants, and if one spot of mold occurs in a propagating bed and is not checked at once it soon spreads over a large space and destroys every cutting or seedling within its reach, but this is not the case with flock in a mushroom bed. If one mushroom is affected with flock every mushroom produced from that piece of spawn is affected, but not one mushroom produced from the pieces of spawn inserted next to this one is affected by it; not even if the mycelium from these several lumps of spawn forms an interlacing web. If the flock is confined to the mushrooms produced from a certain bit of spawn some may ask, will the other pieces of spawn broken from the same brick produce flock-infested mushrooms? No. I have given this point particular attention, [Pg 134] have kept the pieces of each brick close together, and where flock has appeared I have failed to find that the other pieces of spawn from that brick are more liable to produce flock-infested mushrooms than are the pieces of the bricks that, as yet, have not shown any sign of diseased produce.

How general is this disease? In a bed say three feet wide by thirty feet long and of two months' bearing one may get a few as five or as many as fifty flocky mushrooms; one or two may occur to-day, and we may not find another for a week or two, when we may get a whole clump of them, and so on. It is not the large number of them that makes them dangerous, for they never appear in quantity. They sometimes appear among the earliest mushrooms in the bed, but generally not until after the bed has been in bearing condition for a week or two.

What conditions are most favorable or unfavorable to the growth of this disease I do not know; but it is certainly not caused by debility in the mushroom itself, as the parasite attacks healthy, robust mushrooms and debilitated ones indiscriminately. This flocky condition is caused by one or more saprophytic and parasitic fungi of lowly origin, whose various parts are reduced to mere threads, simple or branched, and divided into tubular cells at intervals, or else they are long, continuous microscopic tubes without any partitions, except at those occasional

points where a branch, destined to produce spores, is given off. Generally two or more species of these thread-fungi are present at the same time on the mushroom host, and by the multiplied crossing and interweaving of their threads and branches produce, through their great numbers, the whitish, felted mass of "flock"; while as individuals the threads are so minute as to be scarcely or not at all visible to the naked eye. Similar thread-fungi may often be found in the woods among damp [Pg 135] leaves, under rotten logs, and on those porous fungi which project, shelf-like, from the trunks of trees. At present there is no way known for destroying the "flock," except to take up and destroy every clump of mushrooms attacked by it. Fortunately the disease is not very serious if proper precautions are observed; for, in our own cellars, where mushrooms have been grown year after year for the past eleven years, we get but few flock mushrooms in anybody's bearing. The disease is not more common to-day than it was in any former year. But we give our cellars a thorough cleaning every summer.

Cleaning the Mushroom Houses.—After the season's cropping is finished the mushroom houses and cellars should be thoroughly cleaned. Clear out the old beds, and bring outside all the movable floor and shelf boards, scrape up every bit of loose litter or dirt in the place and throw it out, broom down the walls and whatever boarding is left. Whitewash the walls with hot lime wash, and paint every bit of woodwork liberally with crude oil or kerosene. This is to destroy quillwax and other insect and fungus parasites. If you wish to use again the boards brought outside, broom them over and paint them copiously with kerosene. And if your cellar or house has a dirt floor, a heavy sprinkling of very caustic lime water all over it will do good in ridding it of vermin.

MUSHROOM CULTIVATION-17MBU601A

POSSIBLE QUESTIONS

1. Write about mother spawn.
2. What is mushroom Nutraceuticals?
3. How to preserve the mushroom?
4. What are the different parts of a typical mushroom?
5. What is milky mushroom?
6. Explain the processing and preservation of mushrooms.
7. How to control pests, weed and nematodes during mushroom cultivation?
8. Write about the antibacterial & antifungal activity of mushroom.

SI NO	unit-5	Option A	Option B	Option C	Option D	Answer
1	Dry bubble is characterized by	Puff muddy mushroom	Green mould	Sunken sports on the cap	none of the above	
2	Verticillium chiefly is caused agent for	Dry bubble	False truffle	Mildew	Wet bubble	Dry bubble
3	Example for costing soil and growing mushroom	Fusarium	Aspergillus	Pencillium	Pezizastracoderma	Pezizastracoderma
4	Biotic disease causing agent is	Air pollution	waterpollution	microorganisms	lowtemperature	microorganisms
5	The infected beds usually have a peculiar disagreeable odour, find out the causative agent	Diehliomycesmicrosporus	Fusarium	Pleurotus sp	Pezizastracoderma	Diehliomycesmicrosporus
6	Wet bubble disease was first described in the year of	1435	1788	1887	1888	1888
7	Wet bubble disease is causative agent	Mycogenperniciosa	pink mushroom	white mushroom	Hypomyces rosella	Mycogenperniciosa
8	How to treat the brown plaster disease	Use disinfect	Grown under the low temperature	Change the pH	none of the above	Color changes to cherry red on spore formation
9	Symptoms for lipstick mushroom	mycelium dark	cap turn brown	Thin caps	Color changes to cherry red on spore formation	Color changes to cherry red on spore formation
10	Bacterial blotch is discovered by	Fletcher	Tolaas	Koch	T.J.Burrill	Tolaas
11	Causative agent for bacterial blotch	<i>E.coli</i>	<i>Pseudomonas gingeri</i>	<i>Sallmonella</i>	<i>Pseudomonas tolaasii</i>	<i>Pseudomonas tolaasii</i>
12	Drippy gall disease is observed by	Fletcher	Tolaas	Koch	T.J.Burrill	Fletcher
13	<i>Pseudomonas gingeri</i> cause	Bacterial blotch	mummy disease	fungal disease	ginger blotch	ginger blotch
14	Oyster mushroom is highly affected by	<i>E.coli</i>	<i>Pseudomonas</i>	<i>Pleurotus</i>	<i>Sallmonella</i>	<i>Pleurotus sp</i>

			<i>tolaasii</i>	<i>sp</i>		
15	Mummy disease was first reported in	1942	1947	1950	1948	1942
16	Mummy disease was first reported in the year	Tucker	routeini	Panei	both a and b	both a and b
17	Mummy disease was first reported in	UK	UAE	US	none of the above	UK
18	Drippy Gall disease was first reported in	UK and Netherland	UAE	US	none of the above	UK and Netherland
19	Bacterial Rot disease was first reported in	West Bengal	Tamilnadu	Andhrapra desh	Uttarpradesh	West Bengal
20	_____ flies are also called as mushroom flies.	Sciarid	black gnot	small	none of the above	sciarid
21	_____ are rarely identified from the fly stage.	cecid	Sciarid	black gnot	small	cecid
22	Poison balting flies are present during the _____ period.	harvesting	cropping	picking	casing	cropping
23	After seven days of spawning, _____ should be sprayed on beds.	malathion	methanol	formalin	ethanol	malathion
24	After spraying the malathion,mushroom house should be closed for _____ hours.	1	2	3	4	2
25	_____ spraying on beds should be avoided.	indirect	direct	alternative	none of the above	direct
26	An interval of _____ hours between spraying and picking of mushrooms must be observed.	24	48	72	82	48
27	_____ traps should be used to monitor the number of flies in the spawn running room.	sticky	light	dark	none of the above	sticky
28	_____ are wingless pests.	spring tails	cecid	Sciarid	black gnot	spring tails
29	Springtails are usually found at the base of the _____.	stem	root	cap	none of the above	stem
30	Three species of beetles have been reported to cause damage to _____ mushroom.	Oyster	milky	paddy straw	all of the above	oyster
31	There are more than _____ species of mites which have been reported on mushroom	24	54	44	84	54

32	Nematode contamination in a _____ farm may occur at any time from composting to cropping.	Mushroom	dairy	poultry	both a and b	Mushroom
33	Fresh mushrooms are usually packed in poly packs of less than _____ gauge thickness	100	200	300	400	100
34	Generally, mushroom packs measuring _____ g should be made for retail sale.	200	450	300	both a and b	both a and b
35	_____ is the most popular method of preserving the mushrooms in long term	canning	polythene bags	bottles	none of the above	canning
36	_____ is the most popular method of preserving the mushrooms in short term.	canning	polythene bags	bottles	steeping	steeping
37	_____ is the most popular method of preserving the white button mushrooms.	canning	polythene bags	bottles	none of the above	canning
38	_____ is another method of preserving the mushrooms	canning	polythene bags	steeping	freeze drying	freeze drying
39	_____ is the most popular method of preserving the oyster and paddy straw	canning	polythene bags	steeping	drying	drying
40	Mushroom pickles can be preserved up to	4-3 months	1-2months	3-4months	6-12months	6-12months
41	what is the period between two flushes of mushroo crop?	3-4 days	1-2 months	7-10 days	not certain	7-10 days
42	The optimum temperature required for agaricus biosporus	10-15°c	20-35°c	25-30°c	40-55°c	25-30°c
43	One of the best edible species mushrooms under cultivation is	Saihwai	kanpur	Jhang	kasur	Saihwai
44	Annular ring is absent in mushroom	milky white	paddy straw	button	oyster	button
45	The white button mushroom has been cultivated for its	plant consisting of fine green threads	edible fruit bodies	spores	not seeds	edible fruit bodies



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CLASS:III B.Sc MB

KARPAGAM ACADEMY OF HIGHER EDUCATION

COURSE NAME: MUSHROOM CULTIVATION

COURSE CODE: 17MBU601A

UNIT 5

BATCH-2017-2020

KARHEE