## **Research Paper**

## Degradation of benzo[a]pyrene by *Pleurotus ostreatus* PO-3 in the presence of defined fungal and bacterial co-cultures

Sourav Bhattacharya<sup>1,2</sup>, Arijit Das<sup>1,2</sup>, Muthusamy Palaniswamy<sup>2</sup> and Jayaraman Angayarkanni<sup>3</sup>

<sup>1</sup> Department of Microbiology, Center for Post Graduate Studies, Jain University, Bangalore, Karnataka, India

<sup>2</sup> Department of Microbiology, Karpagam University, Coimbatore, Tamil Nadu, India

<sup>3</sup> Department of Microbial Biotechnology, Bharathiar University, Coimbatore, Tamil Nadu, India

Benzo[a]pyrene, a high molecular weight polycyclic aromatic hydrocarbon possesses carcinogenic, teratogenic, and mutagenic properties. The present study focuses on benzo[a]pyrene degradation by Pleurotus ostreatus PO-3, characterization and identification of metabolites produced and the extent of degradation in the presence of axenic culture of P. ostreatus PO-3 and defined co-cultures of the basidiomycete with bacteria and non-basidiomycete fungi. Thin-layer chromatography revealed that P. ostreatus PO-3 transformed benzo[a]pyrene to polar metabolites. Following degradation, appearance of numerous peaks in the mass spectrum indicated that benzo[a]pyrene degradation was a result of the metabolic activity of P. ostreatus PO-3. A degradation product corresponding to the m/z 284.2 was detected which could possibly be BaPquinone, resulting from the oxidation of benzo[a]pyrene. Compared to the axenic culture of P. ostreatus PO-3 (64.3%), co-cultures of P. ostreatus PO-3 and Penicillium chrysogenum MTCC 787 and P. ostreatus PO-3 and Pseudomonas aeruginosa MTCC 1688 could degrade 86.1 and 75.1% of benzo[a]pyrene, respectively. Thus it could be inferred from the present investigation that the combined catabolic activities of P. ostreatus PO-3 with bacteria and non-basidiomycete fungi can produce synergistic effects to enhance BaP degradation. The increase in the generation of polar metabolites as degradation products from the recalcitrant parent compound advocates the potential application of *P. ostreatus* PO-3 in benzo[*a*]pyrene bioremediation.

**Abbreviations:** BaP – benzo[*a*]pyrene; HMW PAH – high molecular weight polycyclic aromatic hydrocarbon; HPLC – high performance liquid chromatography; m/z – mass-to-charge ratio; TLC – thin layer chromatography

Keywords: Pleurotus ostreatus PO-3 / Benzo[a]pyrene / BaP-quinone / Co-cultures / Bioremediation

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## Introduction

High molecular weight polycyclic aromatic hydrocarbon (HMW PAH) such as benzo[*a*]pyrene (B*a*P) has been listed by the United States Environmental Protection Agency (USEPA) as a priority pollutant owing to its recalcitrant nature. It also behaves as a potent teratogen, mutagen, and carcinogen. Utilization of B*a*P is affected by its low water solubility, resulting in decreased availability for microbial degradation [1]. Several white-rot fungi such as Phanerochaete chrysosporium, Trametes versicolor, Stropharia coronilla, Pleurotus ostreatus, Irpex lacteus, Bjerkandera adusta, etc. can rapidly oxidize BaP with their extracellular lignin modifying enzymes such as lignin peroxidases, manganese peroxidases, laccases, and other oxidases thereby raising interest in the use of these basidiomycetes for the degradation of BaP. However, the inability to isolate a single potent microbial culture showing complete BaP mineralization indicates that such a process in nature is mediated by the cooperative metabolic activities of mixed microbial populations [2].

While BaP degradation by various white-rot fungi is well documented, there is considerably lesser knowledge

**Correspondence:** Sourav Bhattacharya, Department of Microbiology, Center for Post Graduate Studies, Jain University, 18/3, 9th Main, Jayanagar, 3rd Block, Bangalore-560011, Karnataka, India **E-mail:** sourav3011@rediffmail.com